

(10)



Europäisches Patentamt
European Patent Office
Office européen des brevets



(11) Publication number:

0 525 384 A2

(12)

EUROPEAN PATENT APPLICATION(21) Application number: **92110627.4**

(51) Int. Cl.⁵ **C12N 15/60, C12N 9/88,
C12N 15/82, C12N 1/21,
C12N 5/10, A01H 5/00,
A01H 1/02**

(22) Date of filing: **24.06.92**(30) Priority: **31.07.91 US 737851**

(32) Date of publication of application
03.02.93 Bulletin 93/05

(36) Designated Contracting States:
**AT BE CH DE DK ES FR GB GR IT LI LU NL PT
SE**

(71) Applicant: **AMERICAN CYANAMID COMPANY**
1937 West Main Street P.O. Box 60
Stamford Connecticut 06904-0060(US)

(72) Inventor: **Dietrich, Gabriele Elfriede**
11 Merritt Lane
Rocky Hill, New Jersey 08543(US)

(74) Representative: **Wächtershäuser, Günter, Dr.**
Tal 29
W-8000 München 2(DE)

(54) **Imidazolinone resistant ahas mutants.**

(57) The present invention relates to monocot genes encoding a mutant AHAS enzyme that is specifically resistant to imidazolinone herbicides. Exemplary of these genes are corn DNA sequences which encode an amino acid substitution at position 621 of the wild-type AHAS enzyme. The mutant gene can be used to transform other plants to herbicide resistance; in this regard, the invention also provides host cells and vectors containing the gene, which cells and vectors are useful in the transformation process.

EP 0 525 384 A2**BEST AVAILABLE COPY**

This invention relates to novel DNA sequences that encode novel variant forms of acetohydroxy acid synthase enzyme (hereinafter AHAS). The AHAS enzyme is a critical enzyme routinely produced in a variety of plants and a broad range of microorganisms. Normal AHAS function is inhibited by imidazolinone herbicides; however, new AHAS enzymes encoded by the mutant DNA sequences function normally catalytically even in the presence of imidazolinone herbicides and, therefore, confer herbicide resistance upon the plant or microorganism containing them.

The novel DNA sequences are derived from corn and have a substitution of an amino acid at position 621 of the normal AHAS sequence. This substitution in the AHAS gene sequence results in a fully functional enzyme, but renders the enzyme specifically resistant to inhibition by a variety of imidazolinone herbicides. The availability of these variant sequences provides a tool for transformation of different crop plants to imidazolinone herbicide resistance, as well as providing novel selectable markers for use in other types of genetic transformation experiments.

BACKGROUND OF THE INVENTION

The use of herbicides in agriculture is now widespread. Although there are a large number of available compounds which effectively destroy weeds, not all herbicides are capable of selectively targeting the undesirable plants over crop plants, as well as being non-toxic to animals. Often, it is necessary to settle for compounds which are simply less toxic to crop plants than to weeds. In order to overcome this problem, development of herbicide resistant crop plants has become a major focus of agricultural research.

An important aspect of development of herbicide-resistance is an understanding of the herbicide target, and then manipulating the affected biochemical pathway in the crop plant so that the inhibitory effect is avoided while the plant retains normal biological function. One of the first discoveries of the biochemical mechanism of herbicides related to a series of structurally unrelated herbicide compounds, the imidazolinones, the sulfonylureas and the triazopyrimidines. It is now known (Shaner et al, Plant Physiol. 76: 545-546, 1984; U.S. Patent No. 4,761,373) that each of these herbicides inhibits plant growth by interference with an essential enzyme required for plant growth, acetohydroxyacid synthase (AHAS; also referred to as acetolactate synthase, or ALS). AHAS is required for the synthesis of the amino acids isoleucine, leucine and valine.

The AHAS enzyme is known to be present throughout higher plants, as well as being found in a variety of microorganisms, such as the yeast *Saccharomyces cerevisiae*, and the enteric bacteria, *Escherichia coli* and *Salmonella typhimurium*. The genetic basis for the production of normal AHAS in a number of these species has also been well characterized. For example, in both *E. coli* and *S. typhimurium* three isozymes of AHAS exist; two of these are sensitive to herbicides while a third is not. Each of these isozymes possesses one large and one small protein subunit, and map to the *ilvH*, *ilvGM* and *ilvBN* operons. In yeast, the single AHAS isozyme has been mapped to the *ILV2* locus. In each case, sensitive and resistant forms have been identified and sequences of the various alleles have been determined (Friden et al., Nucl. Acid Res. 13: 3979-3993, 1985; Lawther et al., PNAS USA 78: 922-928, 1982; Squires et al., Nucl. Acids Res 11: 5299-5313, 1983; Wek et al, Nucl. Acids Res 13: 4011-4027, 1985; Falco and Dumas, Genetics 109, 21-35, 1985; Falco et al, Nucl. Acids Res 13: 4011-4027, 1985).

In tobacco, AHAS function is encoded by two unlinked genes, *SuRA* and *SuRB*. There is substantial identity between the two genes, both at the nucleotide level and amino acid level in the mature protein, although the N-terminal, putative transit region differs more substantially (Lee et al, EMBO J. 7: 1241-1248, 1988). *Arabidopsis*, on the other hand, has a single AHAS gene, which has also been completely sequenced (Mazur et al., Plant Physiol. 85:1110-1117, 1987). Comparisons among sequences of the AHAS genes in higher plants indicates a high level of conservation of certain regions of the sequence; specifically, there are at least 10 regions of sequence conservation. It has previously been assumed that these conserved regions are critical to the function of the enzyme, and that retention of that function is dependent upon substantial sequence conservation.

It has been recently reported (U.S. Patent No. 5,013,659) that mutants exhibiting herbicide resistance possess mutations in at least one amino acid in one or more of these conserved regions. In particular, substitution of certain amino acids for the wild type amino acid at these specific sites in the AHAS protein sequence have been shown to be tolerated, and indeed result in herbicide resistance of the plant possessing this mutation, while retaining catalytic function. The mutations described therein encode either cross resistance for imidazolinones and sulfonylureas or sulfonylurea-specific resistance, but no imidazolinone-specific mutations were disclosed. These mutations have been shown to occur at both the *SuRA* and *SuRB* loci in tobacco; similar mutations have been isolated in *Arabidopsis* and yeast.

Imidazolinone-specific resistance has been reported elsewhere in a number of plants. U.S. Patent No.

4,761,373 generally described an altered AHAS as a basis of herbicide resistance in plants, and specifically disclosed certain imidazolinone resistant corn lines. Haughn et al. (Mol. Gen. Genet. 211:266-271, 1988) disclosed the occurrence of a similar phenotype in *Arabidopsis*. Sathasivan et al. (Nucl. Acid Res. 18:2188, 1990) identified the imidazolinone-specific resistance in *Arabidopsis* as being based on a mutation at position 653 in the normal AHAS sequence. In accordance with the present invention, a gene encoding imidazolinone-specific resistance in a monocot has now been isolated and determined to be associated with a single amino acid substitution in a wild-type monocot AHAS amino acid sequence.

SUMMARY OF THE INVENTION

The present invention provides novel nucleic acid sequences encoding functional monocot AHAS enzymes insensitive to imidazolinone herbicides. The sequences in question comprise a mutation in the codon encoding the amino acid serine at position 621 in the corn (maize) AHAS sequence, or in the corresponding position in other monocot sequences. Other monocots, such as wheat, are also known to exhibit imidazolinone specific mutations (e.g., ATCC Nos. 40994-97). In corn, the wild type sequence has a serine at this position. In a preferred embodiment, the substitution is asparagine for serine, but alternate substitutions for serine include aspartic acid, glutamic acid, glutamine and tryptophane. Although the claimed sequences are originally derived from monocots, the novel sequences are useful in methods for producing imidazolinone resistant cells in any type of plant, said methods comprising transforming a target plant cell with one or more of the altered sequences provided herein. Alternatively, mutagenesis is utilized to create mutants in plant cells or seeds containing a nucleic acid sequence encoding an imidazolinone insensitive AHAS. In the case of mutant plant cells isolated in tissue culture, plants which possess the imidazolinone resistant or insensitive trait are then regenerated. The invention thus also encompasses plant cells, bacterial cells, fungal cells, plant tissue cultures, adult plants, and plant seeds that possess a mutant nucleic acid sequence and which express functional imidazolinone resistant AHAS enzymes.

The availability of these novel herbicide resistant plants enables new methods of growing crop plants in the presence of imidazolinones. Instead of growing non-resistant plants, fields may be planted with the resistant plants produced by mutation or by transformation with the mutant sequences of the present invention, and the field routinely treated with imidazolinones, with no resulting damage to crop plants.

The mutant nucleic acids of the present invention also provide novel selectable markers for use in transformation experiments. The nucleic acid sequence encoding a resistant AHAS is linked to a second gene prior to transfer to a host cell, and the entire construct transformed into the host. Putative transformed cells are then grown in culture in the presence of inhibitory amounts of herbicide; surviving cells will have a high probability of having successfully acquired the second gene of interest. Alternately, the resistant AHAS gene can be cotransformed on an independent plasmid with the gene of interest, whereby about 50% of all transformants can be expected to have received both genes.

The following definitions should be understood to apply throughout the specification and claims. A "functional" or "normal" AHAS enzyme is one which is capable of catalyzing the first step in the pathway for synthesis of the essential amino acids isoleucine, leucine and valine. A "wild-type" AHAS sequence is a sequence present in an imidazolinone sensitive member of a given species. A "resistant" plant is one which produces a mutant but functional AHAS enzyme, and which is capable of reaching maturity when grown in the presence of normally inhibitory levels of imidazolinone. The term "resistant", as used herein, is also intended to encompass "tolerant" plants, i.e., those plants which phenotypically evidence adverse, but not lethal, reactions to the imidazolinone.

BRIEF DESCRIPTION OF THE FIGURES

Figure 1: AHAS enzyme activity in 10-day old maize seedlings (corn lines A619 or X112) in the presence of imazethapyr (Pursuit™ A) or chlorsulfuron (Oust™ B). Herbicide resistant AHAS activity is calculated as percentage of AHAS activity in the absence of inhibitor. The standard error between experiments is 10%.

Figure 2: Southern analysis of genomic clones in phage EMBL3. Phages 12-1A (from W22), 12-7A, 18-8A, 12-11, and 12-17A (From X112) are digested with XbaI or SalI, separated on a 1% agarose gel, transferred onto nitrocellulose and hybridized with an AHAS cDNA fragment as probe.

Figure 3: Southern analysis of genomic DNA from corn lines X112, XA17, QJ22, A188 and B73. The DNA is digested with XbaI, separated on a 1% agarose gel, transferred onto nitrocellulose and hybridized with an AHAS cDNA fragment as probe.

Figure 4: Restriction map of plasmid pCD8A. The mutant AHAS gene from X112 was subcloned as a

8kb PstI fragment into vector pKS(+). The location and orientation of the AHAS gene is indicated by an arrow. The restriction sites of PstI, XhoI, HindIII, XbaI and ClaI are represented by symbols.

Figure 5: Nucleotide sequencing gel of the non-coding strand (A) and the double stranded DNA sequence (B) of AHAS clones W22/4-4, B73/10-4 and X112/8A in the region of amino acids 814 to 833. The position of the cytosine to thymidine transition is indicated by an arrow.

Figure 6: Nucleotide and deduced amino acid sequences of the X112/8A mutant AHAS gene.

Figure 7: Nucleotide sequence alignment of X112/8A, B73/7-4 and W22/1A *als2* genes. (*) marks the base change causing the mutation at position 621, (#) differences from the B73/7-4 sequence and (>) represents silent changes.

Figure 8: Amino acid sequences and alignment of X112/8A, B73/7-4 and W22/1A *als2* genes. (*) marks the mutation at position 621, (#) marks differences from the B73/7-4 sequence, and (>) represents silent changes.

DETAILED DESCRIPTION OF THE INVENTION

The gene of the present invention is isolatable from corn maize line X112 (seed deposited with the American Type Culture Collection as Accession Number 75051), and has been inserted into plasmid pX112/8A (deposited with the American Type Culture Collection as Accession Number 68843); it is also isolatable from any other imidazolinone-specific herbicide resistant mutant, such as the corn line QJ22 (deposited as a cell culture with the American Type Culture Collection as Accession Number 40129), or the various wheat plants (seed deposited with the American Type Culture Collection as Accession Numbers 40994, 40995, 40996, or 40997). A genomic DNA library is created, for example, in phage ENBL-3 with DNA from one of the imidazolinone resistant mutants, preferably one which is homozygous for the resistance trait, and is screened with a nucleic acid probe comprising all or a part of a wild-type AHAS sequence.

In maize, the AHAS gene is found at two loci, *als1* and *als2* (Burr and Burr, Trends in Genetics 7:55-61, 1991); the homology between the two loci is 95% at the nucleotide level. The mutation in X112 is mapped to locus *als2* on chromosome 5, whereas the nonmutant gene is mapped to locus *als1* on chromosome 4 (Newhouse et al., "Imidazolinone-resistant crops". In The Imidazolinone Herbicides, Shaner and O'Connor (Eds.), CRC Press, Boca Raton, FL, in Press). Southern analysis identifies some clones containing the mutant *als2* gene, and some containing the non-mutant *als1* gene. Both types are subcloned into sequencing vectors, and sequenced by the dideoxy sequencing method.

Sequencing and comparison of wild type and mutant AHAS genes shows a difference of a single nucleotide in the codon encoding the amino acid at position 621 (Figure 5). Specifically, the codon AGT encoding serine in the wild type is changed to AAT encoding asparagine in the mutant AHAS (Figure 8). The mutant AHAS gene is otherwise similar to the wild type gene, encoding a protein having 838 amino acids, the first 40 of which constitute a transit peptide which is thought to be cleaved during transport into the chloroplast in vivo. The sequence of the *als1* non-mutant gene from X112 appears to be identical to the *als1* gene from B73.

The mutant genes of the present invention confer resistance to imidazolinone herbicides, but not to sulfonylurea herbicides. Types of herbicides to which resistance is conferred are described for example in U.S. Patent Nos. 4,188,487; 4,201,565; 4,221,586; 4,297,128; 4,554,013; 4,608,079; 4,638,068; 4,747,301; 4,850,514; 4,698,092; 4,701,208; 4,709,036; 4,752,323; 4,772,311; and 4,798,619.

It will be understood by those skilled in the art that the nucleic acid sequence depicted in Figure 6 is not the only sequence which can be used to confer imidazolinone-specific resistance. Also contemplated are those nucleic acid sequences which encode an identical protein but which, because of the degeneracy of the genetic code, possess a different nucleotide sequence. The invention also encompasses genes encoding AHAS sequences in which the aforementioned mutation is present, but which also encode one or more silent amino acid changes in portions of the molecule not involved with resistance or catalytic function. Also contemplated are gene sequences from other imidazolinone resistant monocots which have a mutation in the corresponding region of the sequences.

For example, alterations in the gene sequence which result in the production of a chemically equivalent amino acid at a given site are contemplated; thus, a codon for the amino acid alanine, a hydrophobic amino acid, can readily be substituted by a codon encoding another hydrophobic residue, such as glycine, or may be substituted with a more hydrophobic residue such as valine, leucine, or isoleucine. Similarly, changes which result in substitution of one negatively charged residue for another, such as aspartic acid for glutamic acid, or one positively charged residue for another, such as lysine for arginine, can also be expected to produce a biologically equivalent product. The invention also encompasses chimeric genes, in which the substituted portion of the corn AHAS gene is recombined with unaltered portions of the normal AHAS genes

from other species. Thus, throughout the specification and claims, wherever the term "imidazolinone-specific resistant corn AHAS gene" is used, it is intended to cover each of these alternate embodiments as well as the sequence of Figure 6.

Isolated AHAS DNA sequences of the present invention are useful to transform target crop plants, and thereby confer imidazolinone resistance. A broad range of techniques currently exist for achieving direct or indirect transformation of higher plants with exogenous DNA, and any method by which the novel sequence can be incorporated into the host genome, and stably inherited by its progeny, is contemplated by the present invention. The imidazolinone specific resistance trait is inherited as a single dominant nuclear gene. The level of imidazolinone resistance is increased when the gene is present in a homozygous state; such corn plants, for example, have a resistance level of about 1,000 times that of a non-resistant plant. Plants heterozygous for the trait, however, have a resistance of about 50-500 times that of a non-resistant plant.

Transformation of plant cells can be mediated by the use of vectors. A common method of achieving transformation is the use of *Agrobacterium tumefaciens* to introduce a foreign gene into the target plant cell. For example, the mutant AHAS sequence is inserted into a plasmid vector containing the flanking sequences in the Ti-plasmid T-DNA. The plasmid is then transformed into *E. coli*. A triparental mating among this strain, an *Agrobacterium* strain containing a disarmed Ti-plasmid containing the virulence functions needed to effect transfer of the AHAS containing T-DNA sequences into the target plant chromosome, and a second *E. coli* strain containing a plasmid having sequences necessary to mobilize transfer of the AHAS construct from *E. coli* to *Agrobacterium* is carried out. A recombinant *Agrobacterium* strain, containing the necessary sequences for plant transformation is used to infect leaf discs. Discs are grown on selection media and successfully transformed regenerants are identified. The recovered plants are resistant to the effects of herbicide when grown in its presence. Plant viruses also provide a possible means for transfer of exogenous DNA.

Direct uptake of plant cells can also be employed. Typically, protoplasts of the target plant are placed in culture in the presence of the DNA to be transferred, and an agent which promotes the uptake of DNA by protoplast. Useful agents in this regard are polyethylene glycol or calcium phosphate.

Alternatively, DNA uptake can be stimulated by electroporation. In this method, an electrical pulse is used to open temporary pores in a protoplast cell membrane, and DNA in the surrounding solution is then drawn into the cell through the pores. Similarly, microinjection can be employed to deliver the DNA directly into a cell, and preferably directly into the nucleus of the cell.

In each of the foregoing techniques, transformation occurs in a plant cell in culture. Subsequent to the transformation event, plant cells must be regenerated to whole plants. Techniques for the regeneration of mature plants from callus or protoplast culture are now well known for a large number of different species (see, e.g., Handbook of Plant Cell Culture, Vols. 1-5, 1983-1989 McMillan, N.Y.) Thus, once transformation has been achieved, it is within the knowledge in the art to regenerate mature plants from the transformed plant cells.

Alternate methods are also now available which do not necessarily require the use of isolated cells, and therefore, plant regeneration techniques, to achieve transformation. These are generally referred to as "ballistic" or "particle acceleration" methods, in which DNA coated metal particles are propelled into plant cells by either a gunpowder charge (Klein et al., Nature 327: 70-73, 1987) or electrical discharge (EPO 270 356). In this manner, plant cells in culture or plant reproductive organs or cells, e.g. pollen, can be stably transformed with the DNA sequence of interest.

In certain dicots and monocots direct uptake of DNA is the preferred method of transformation. For example, in corn, the cell wall of cultured cells is digested in a buffer with one or more cell wall degrading enzymes, such as cellulase, hemicellulase and pectinase, to isolate viable protoplasts. The protoplasts are washed several times to remove the enzymes, and mixed with a plasmid vector containing the gene of interest. The cells can be transformed with either PEG (e.g. 20% PEG 4000) or by electroporation. The protoplasts are placed on a nitrocellulose filter and cultured on a medium with embedded corn cells functioning as feeder cultures. After 2-4 weeks, the cultures in the nitrocellulose filter are placed on a medium containing about 0.32 μ M of the imidazolinone and maintained in the medium for 1-2 months. The nitrocellulose filters with the plant cells are transferred to fresh medium with herbicides and nurse cells every two weeks. The untransformed cells cease growing and die after a few weeks.

The present invention can be applied to transformation of virtually any type of plant, both monocot and dicot. Among the crop plants for which transformation to herbicide resistance is contemplated are corn, wheat, rice, millet, oat, barley, sorghum, sunflower, sweet potato, alfalfa, sugar beet, Brassica species, tomato, pepper, soybean, tobacco, melon, squash, potato, pea, cotton, or cacao. The novel sequences may also be used to transform ornamental species, such as rose, and woody species, such as pine and poplar.

The novel sequences of the invention also are useful as selectable markers in plant genetics studies. For example, in plant transformation, sequences encoding imidazolinone resistance could be linked to a gene of interest which is to be used to transform a target imidazolinone sensitive plant cell. The construct comprising both the gene of interest and the imidazolinone resistant sequence are introduced into the plant cell, and the plant cells are then grown in the presence of an inhibitory amount of an imidazolinone. Alternately, the second gene of interest can be cotransformed, on a separate plasmid, into the host cells. Plant cells surviving such treatment presumably have acquired the resistance gene as well as the gene of interest, and therefore, only transformants survive the selection process with the herbicide. Confirmation of successful transformation and expression of both genes can be achieved by Southern hybridization of genomic DNA, by PCR or by observation of the phenotypic expression of the genes.

The invention is further illustrated by the following non-limiting examples.

EXAMPLES

1. Confirmation of Whole Plant Herbicide Resistance in XI12

XI12 plants are treated with herbicides at 10 days to the V3 leaf stage (4-5 leaves, of which 3 have visible ligules). Imazethapyr is applied at rates of 2000, 500, 250, 125 and 62.5 g/ha. Chlorsulfuron is applied at 32, 16, 8, 4 and 2 g/ha. Plants are treated postemergence at a spray volume of 400 l/ha. After spraying, plants are placed in the greenhouse for further observation.

XI12 plants are unaffected at all rates of imazethapyr treatment; however, no visible increased resistance to chlorsulfuron is noted. Thus, XI12 displays selective resistance to the imidazolinone at the whole plant level (See Figure 1).

The resistance in XI12 is also shown to be inherited as a single dominant allele of a nuclear gene. Heterozygous resistant XI12 are selfed, and the selfed progeny segregate in the 3 resistant:1 susceptible ratio expected for a single dominant allele of a nuclear gene. In this study, the segregating seedlings are sprayed postemergence with lethal doses of imazethapyr (125 or 250 g/ha) following spraying protocols described above, to establish segregation for resistance.

2. AHAS Extraction

Seeds of XI12 are sown in soil in a greenhouse maintained at day/night temperature of 80°C and 15 hour photoperiod. Plants are harvested two weeks after planting. The basal portion of the shoot is used for the extraction of AHAS. 5 g of the tissue is powdered in liquid nitrogen and then homogenized in AHAS assay buffer comprising 100 mM potassium phosphate buffer (pH 7.5) containing 10 mM pyruvate, 5 mM MgCl₂, 5 mM EDTA, 100 µM FAD (flavin adenine dinucleotide), 1 mM valine, 1 mM leucine, 10% glycerol and 10 mM cysteine. The homogenate is centrifuged at 10,000 rpm for 10 minutes and 3 ml of the supernatant are applied onto an equilibrated Bio-Rad Econo-Desalting column (10 DG) and eluted with 4 ml AHAS assay buffer.

AHAS activity is measured by estimation of the product, acetolactate, after conversion by decarboxylation in the presence of acid to acetoin. Standard reaction mixtures contain the enzyme in 50 mM potassium phosphate (pH 7.0) containing 100 mM sodium pyruvate, 10 mM MgCl₂, 1 mM thiamine pyrophosphate, 10 µM FAD, and appropriate concentrations of different inhibitors. This mixture is incubated at 37°C for 1 to 3 hours depending upon the experiment. At the end of this incubation period, the reaction is stopped with the addition of H₂SO₄ to make a final concentration of 0.85% H₂SO₄ in the tube. The reaction product is allowed to decarboxylate at 60°C for 15 minutes. The acetoin formed is determined by incubating with creatine (0.17%) and 1-naphthol (1.7% in 4N NaOH). The absorption of color complex formed is measured at 520 nm.

AHAS activity from B73, A619, or other wild-type maize lines is highly sensitive to inhibition by imazethapyr (PURSUIT™) with an I₅₀ of 1 µM (See Figure 1). Contrary to this observation, XI12 shows 70-80% of enzyme activity at the highest concentrations (100 µM) of PURSUIT™ or ARSENAL™ (imazepyr), and about 70% in the presence of SCEPTER™ (imazequin). This result shows a 100-fold increase in tolerance of AHAS activity from XI12 to imazethapyr as compared to the in vitro AHAS activity from A619. Sensitivity of AHAS activity from the two lines to sulfonylureas gives a different picture. In the presence of OUST™ (sulfometuron methyl), at 100 nM, AHAS activity of XI12 is only 15-20%. AHAS activity of A619 in the presence of OUST™ is 5-10%, and in the presence of PURSUIT™ is 15-20% (See Figure 1).

3. Cloning of XI12 AHAS Genes

Seeds of the XI12 mutant derived from an Imidazolinone resistant corn tissue culture line are planted; plants obtained therefrom appear to be segregating for the mutant AHAS phenotype. In order to obtain homozygous resistant seed material, a population of XI12 mutant plants are selfed. After selecting for herbicide resistance for three consecutive growing seasons, the seeds are homozygous for the mutant AHAS gene. Homozygous seeds are collected and used to grow seedlings to be used in AHAS gene isolation.

DNA is extracted from 7 days old etiolated seedlings of a homozygous XI12 line. 60 g of plant tissue is powdered in liquid nitrogen, and transferred into 108 ml DNA extraction buffer (1.4 M NaCl, 2.0% Ctab (hexadecyl trimethyl ammonium bromide), 100 mM Tris-Cl pH 8.0, 20 mM EDTA, 2% Mercaptoethanol) and 54 ml water. After incubation at 50-60°C for 30 minutes the suspension is extracted with an equal amount of chloroform. The DNA is precipitated by adding an equal amount of precipitation buffer (1% Ctab, 50 mM Tris-Cl pH 8.0, 10 mM EDTA). To purify the genomic DNA, a high speed centrifugation in 6.6M CsCl and ethidium bromide is performed (Ti80 rotor, 50,000 rpm, 20°C, 24 hours). The purified DNA is extracted with salt saturated Butanol and dialyzed for 25 hours against 3 changes of 1 l dialysis buffer (10 mM Tris-Cl Ph 8.0, 1 mM EDTA, 0.1M NaCl). The concentration of the XI12 genomic DNA is determined spectrophotometrically to be 310 mg/ml. The yield is 0.93 mg.

The XI12 genomic DNA is used to create a genomic library in the phage EMBL-3. The DNA is partially digested with MboI and the fragments are separated on a sucrose gradient to produce size range between 8 to 22 kb before cloning into the BamHI site of EMBL-3. After obtaining 2.1×10^6 independent clones, the library is amplified once. The titer of the library is determined 9×10^{10} pfu/ml.

To obtain probes for analysis of the XI12 library, a W22 (wild-type) cDNA library in lambda gt11, purchased from Clontech Inc., CA, is screened with an 800 nt BamHI probe isolated from Arabidopsis AHAS genomic clone. The phages are plated in a density of 50,000 pfu/15 cm plate, transferred onto nitrocellulose filters, prehybridized in 6x SSC, 0.2% SDS for 2 hours and hybridized with the Arabidopsis AHAS probe in 6x SSC, 0.2% SDS overnight. One positive phage is identified, further purified and used for subcloning of a 1.1 kb EcoRI fragment. The 1.1 kb EcoRI fragment is subcloned into pGemA-4 and used as a probe to identify the XI12 AHAS genes.

The XI12 genomic library is plated on 12 15-cm plates (concentration of 50,000 pfu/plate) and is screened with the W22 AHAS cDNA probe. The filters are prehybridized (2 hours) and hybridized (over night) in Church buffer (0.5 M Na Phosphate, 1 mM EDTA, 1% BSA, 7% SDS) at 65°C and washed at 65°C in 2x SSC, 0.2% SDS and 0.3 x SSC, 0.2% SDS. 12 positive plaques are obtained from a total of 7.5×10^5 pfu screened and 5 positive clones are further purified and isolated according to Chisholm (BioTechniques 7:21-23, 1989). Southern analysis (See Figure 2) showed that the phage clones represented two types of AHAS clones: Type-1 clones contain one large XbaI (>6.5 kb) fragment hybridizing to the AHAS cDNA probe, Type-2 clones contained two 2.7 and 3.7 kb XbaI fragments hybridizing to the AHAS cDNA probe. Genomic Southern of XI12 DNA demonstrated, that the XbaI fragments obtained by digesting genomic DNA and by hybridizing to the AHAS cDNA probe correspond to the XbaI fragments identified in the XI12 phage clones (See Figure 3). Restriction digest and Southern Analysis also demonstrate that of the 5 AHAS clones, one clone represents the mutant als2 genes and four represent the als1 gene.

The AHAS genes corresponding to the mutant locus located on chromosome 5 (clone 12/8A) and the non-mutant locus located on chromosome 4 (clone 12/17A) are subcloned as a PstI fragment (clone 12/8A) or as XbaI fragment (12/17A) into the sequencing vector pBluescript II KSml3(+) (pKS+; Stratagene). Both 2.7 kb and 3.7 kb XbaI fragments from phage 12/17A contain one complete copy of AHAS genes which are identified. The sequence of each is obtained by dideoxy sequencing (Pharmacia T7 sequencing Kits) using primers complementary to the AHAS coding sequence.

The methods of DNA extraction, cloning of the genomic library and screening of the library are as described for the XI12 genomic DNA. The B73 AHAS genes are subcloned into the sequencing vector pKS+ as XbaI fragments and are sequenced. The sequence is obtained by dideoxy sequencing, using primers complementary to the AHAS coding sequence as described for the SI12 AHAS genes.

A W22 genomic library in EMBL3 purchased from Clontech Inc., CA is screened. The phages are plated in a density of 50,000 pfu/15 cm plate, transferred onto nitrocellulose filters, and hybridized with the W22 AHAS cDNA probe described above (prehybridization and hybridization conditions: 6 x SSC, 0.5% SDS, 1x Denhardt's 100 mg/ml calf thymus DNA, 65°C, washing conditions: 3X x SSC, 0.2% SDS for 2 hours at 65°C, and 0.3 x SSC, 0.2% SDS for 2 hours). Two positive phages (12/1A and 12/4-4) are identified and further purified.

The W22 genomic clone 12/1A is subcloned as two 0.78 kb (pGemA-4) and 3.0 kb (pGemA-14; Promega) SalI fragments into the vector pGemA-2, and as a 6.5 kb XbaI fragment into the vector pKS+ (pCD200). The coding strand sequence of the W22 AHAS gene is obtained by dideoxy sequencing of

nested deletions created from subclones pGem A-14 and pGem A-4 of phage 12-1A. This sequence is used to design oligonucleotides complementary to the AHAS non-coding strand. The sequence of the non-coding strand is obtained by dideoxy sequencing of clone pCD200 using primers complementary to the coding strand. Upon complementing the sequencing of the W22 AHAS gene, primers of both DNA strands are designed and used for the sequencing of the AHAS genes isolated from the X112 and B73 genomic libraries.

4. Cloning of QJ22 AHAS Genes

The sequence of the gene encoding AHAS in the maize line QJ22, which is selectively resistant to imidazolinones, is also determined. A genomic library of QJ22 is prepared in an EMBL3 vector. A library of 800,000 phage is screened with an 850 nucleotide SalI/ClaI fragment isolated from an AHAS clone (A-4) derived from the wild-type maize line W22. Five positive phages are picked and submitted to a second round of screening to partially purify the phage. The partially purified phage are analyzed by PCR to determine if any clones represent the QJ22 *als1* gene. Such clones are identified as a 3.7kb XbaI fragment with a gene specific SmaI site at position 495. The second screen indicates the presence of a single positive clone with these characteristics.

The PCR product is purified using a commercial kit (Magic-PCR Preps) from Promega; and the purified DNA is sequenced with a Taq polymerase sequencing system "fmol", also from Promega. Sequence analysis of both strands of the DNA of the QJ22 mutant AHAS shows a nucleotide transition from G to A in the codon for amino acid 621. This mutation is identical to the one seen in X112 and the remainder of the sequence is typical of an *als1* gene.

RESULTS

The sequence of the mutant AHAS genes is compared with the sequences obtained from the wild type corn lines B73 and W22 (See Figure 7). The X112 mutant gene (X112/8A) and the QJ22 mutant gene and the wild type gene are identical except for the amino acid change at position 621, causing a single nucleotide transition from AGT to AAT (See Figure 8). The X112 mutant X112/8A and the wild-type B73/7-4 gene show an additional difference at position 63. On the other hand, the non-mutant X112 AHAS gene cloned in X112/17A is completely homologous to the corresponding B73/10-2 in the region coding for the mature AHAS protein (data not shown).

DEPOSIT OF BIOLOGICAL MATERIALS

The following biological materials were deposited with the American Type Culture Collection, 12301 Parklawn Drive, Rockville, Maryland, 20857, as follows:

E. coli XLI Blue harboring plasmid pX12/8A, deposited on July 3, 1991, Accession Number ATCC 68643

X112 corn seed deposited on July 16, 1991, Accession Number ATCC 75051.

Sequence Listings

5 Sequence ID No.: 1

Sequence Type: Nucleotide and Amino Acid

10 Sequence Length: 1969 BP's and 638 Amino Acids

Strandedness: Single

15 Topology: Linear

20 Original Source Organism: Zea mays

Properties: Herbicide Resistant AHAS Enzyme

25 AACCCCTCGCG CCGCCTCCGA GACAGCCGCC GCAACC 36

30 ATG GCC ACC GCC GCC GCC GCG TCT ACC GCG CTC ACT 72

Met Ala Thr Ala Ala Ala Ser Thr Ala Leu Thr
1 5 10

35 GGC GCC ACT ACC GCT GCG CCC AAG GCG AGG CGC CGG 108

Gly Ala Thr Thr Ala Ala Pro Lys Ala Arg Arg Arg
15 20

40 GCG CAC CTC CTG GCC ACC GCG CGC GCC CTC GCC GCG 144

Ala His Leu Leu Ala Thr Arg Arg Ala Leu Ala Ala
25 30 35

45 CCC ATC AGG TGC TCA GCG GCG TCA CCC GCC ATG CCG 180

50 Pro Ile Arg Cys Ser Ala Ala Ser Pro Ala Met Pro
40 45

55

| | | |
|----|---|-----|
| 5 | ATG GCT CCC CGC GCC ACC CGC CTC CGG CCG TGG GGC | 216 |
| | Met Ala Pro Pro Ala Thr Pro Leu Arg Pro Trp Gly | |
| | 50 55 60 | |
| 10 | CCC ACC GAT CCC CGC AAG GGC GCC GAC ATC CTC GTC | 252 |
| | Pro Thr Asp Pro Arg Lys Gly Ala Asp Ile Leu Val | |
| | 65 70 | |
| 15 | GAG TCC CTC GAG CGC TGC GGC GTC CGC GAC GTC TTC | 288 |
| | Glu Ser Leu Glu Arg Cys Gly Val Arg Asp Val Phe | |
| 20 | 75 80 | |
| | GCC TAC CCC GGC GGC GCG TCC ATG GAG ATC CAC CAG | 324 |
| | Ala Tyr Pro Gly Gly Ala Ser Met Glu Ile His Gln | |
| 25 | 85 90 95 | |
| | GCA CTC ACC CGC TCC CCC GTC ATC GCC AAC CAC CTC | 360 |
| 30 | Ala Leu Thr Arg Ser Pro Val Ile Ala Asn His Leu | |
| | 100 105 | |
| 35 | TTC CGC CAC GAG CAA GGG GAG GCC TTT GCG GCC TCC | 396 |
| | Phe Arg His Glu Gln Gly Glu Ala Phe Ala Ala Ser | |
| | 110 115 120 | |
| 40 | GGC TAC GCG CGC TCC TCG GGC CGC GTC GGC GTC TGC | 432 |
| | Gly Tyr Ala Arg Ser Ser Gly Arg Val Gly Val Cys | |
| | 125 130 | |
| 45 | ATC GCC ACC TCC GGC CCC GGC GCC ACC AAC CTT GTC | 468 |
| | Ile Ala Thr Ser Gly Pro Gly Ala Thr Asn Leu Val | |
| 50 | 135 140 | |
| | TCC GCG CTC GCC GAC GCG CTG CTC GAT TCC GTC CCC | 504 |
| | Ser Ala Leu Ala Asp Ala Leu Leu Asp Ser Val Pro | |
| 55 | 145 150 155 | |

| | | |
|----|---|-----|
| 5 | ATG GTC GCC ATC ACG GGA CAG GTG CCG CGA CGC ATG | 540 |
| | Met Val Ala Ile Thr Gly Gln Val Pro Arg Arg Met | |
| | 160 165 | |
| 10 | ATT GGC ACC GAC GCC TTC CAG GAG ACG CCC ATC GTC | 576 |
| | Ile Gly Thr Asp Ala Phe Gln Glu Thr Pro Ile Val | |
| | 170 175 180 | |
| 15 | GAG GTC ACC CGC TCC ATC ACC AAG CAC AAC TAC CTG | 612 |
| | Glu Val Thr Arg Ser Ile Thr Lys His Asn Tyr Leu | |
| | 185 190 | |
| 20 | GTC CTC GAC GTC GAC GAC ATC CCC CGC GTC GTG CAG | 648 |
| | Val Leu Asp Val Asp Asp Ile Pro Arg Val Val Gln | |
| | 195 200 | |
| 25 | GAG GCT TTC TTC CTC GCC TCC TCT GGT CGA CCG GGG | 684 |
| | Glu Ala Phe Phe Leu Ala Ser Ser Gly Arg Pro Gly | |
| | 205 210 215 | |
| 30 | CCG GTG CTT GTC GAC ATC CCC AAG GAC ATC CAG CAG | 720 |
| | Pro Val Leu Val Asp Ile Pro Lys Asp Ile Gln Gln | |
| | 220 225 | |
| 35 | CAG ATG GCG GTG CCT GTC TGG GAC AAG CCC ATG AGT | 756 |
| | Gln Met Ala Val Pro Val Trp Asp Lys Pro Met Ser | |
| | 230 235 240 | |
| 40 | CTG CCT GGG TAC ATT GCG CGC CTT CCC AAG CCC CCT | 792 |
| | Leu Pro Gly Tyr Ile Ala Arg Leu Pro Lys Pro Pro | |
| | 245 250 | |
| 45 | GCG ACT GAG TTG CTT GAG CAG GTG CTG CGT CTT GTT | 828 |
| | Ala Thr Glu Leu Leu Glu Gln Val Leu Arg Leu Val | |
| | 255 260 | |
| 50 | | |
| 55 | | |

5

10

15

20

30

35

40

45

50

| | | |
|----|---|------|
| | AAG CAG CCA CAT GTG TCC ATC TGT GCA GAT GTT AAG | 1188 |
| | Lys Gln Pro His Val Ser Ile Cys Ala Asp Val Lys | |
| 5 | 375 380 | |
| | CTT GCT TTG CAG GGC ATG AAT GCT CTT CTT GAA GGA | 1224 |
| | Leu Ala Leu Gln Gly Met Asn Ala Leu Leu Glu Gly | |
| 10 | 385 390 395 | |
| | AGC ACA TCA AAG AAG AGC TTT GAC TTT GGC TCA TGG | 1260 |
| 15 | Ser Thr Ser Lys Lys Ser Phe Asp Phe Gly Ser Trp | |
| | 400 405 | |
| | AAC GAT GAG TTG GAT CAG CAG AAG AGG GAA TTC CCC | 1296 |
| 20 | Asn Asp Glu Leu Asp Gln Gln Lys Arg Glu Phe Pro | |
| | 410 415 420 | |
| | CTT GGG TAT AAA ACA TCT AAT GAG GAG ATC CAG CCA | 1332 |
| 25 | Leu Gly Tyr Lys Thr Ser Asn Glu Glu Ile Gln Pro | |
| | 425 430 | |
| 30 | CAA TAT GCT ATT CAG GTT CTT GAT GAG CTG ACG AAA | 1368 |
| | Gln Tyr Ala Ile Gln Val Leu Asp Glu Leu Thr Lys | |
| 35 | 435 440 | |
| | GGC GAG GCC ATC ATC GGC ACA GGT GTT GGG CAG CAC | 1404 |
| 40 | Gly Glu Ala Ile Ile Gly Thr Gly Val Gly Gln His | |
| | 445 450 455 | |
| | CAT ATG TGG GCG GCA CAG TAC TAC ACT TAC AAG CGG | 1440 |
| 45 | Gln Met Trp Ala Ala Gln Tyr Tyr Thr Tyr Lys Arg | |
| | 460 465 | |
| 50 | CCA AGG CAG TGG TTG TCT TCA GCT GGT CTT GGG GCT | 1476 |
| | Pro Arg Gln Trp Leu Ser Ser Ala Gly Leu Gly Ala | |
| | 470 475 480 | |
| 55 | | |

ATG GGA TTT GGT TTG CCG GCT GCT GCT GGT GCT TCT 1512
Met Gly Phe Gly Leu Pro Ala Ala Ala Gly Ala Ser

485

490

GTG GCC AAC CCA GGT GTT ACT GTT GTT GAC ATC GAT 1548
Val Ala Asn Pro Gly Val Thr Val Val Asp Ile Asp

495

500

GGA GAT GGT AGC TTT CTC ATG AAC GTT CAG GAG CTA 1584
Gly Asp Gly Ser Phe Leu Met Asn Val Gln Glu Leu
505 510 515

GCT ATG ATC CGA ATT GAG AAC CTC CCG GTG AAG GTC 1620
Ala Met Ile Arg Ile Glu Asn Leu Pro Val Lys Val
520 525

TTT GTG CTA AAC AAC CAG CAC CTG GGG ATG GTG GTG 1656
Phe Val Leu Asn Asn Gln His Leu Gly Met Val Val
530 535 540

CAG TGG GAG GAC AGG TTC TAT AAG GCC AAC AGA GCG 1692
Gln Trp Glu Asp Arg Phe Tyr Lys Ala Asn Arg Ala
545 550

CAC ACA TAC TTG GGA AAC CCA GAG AAT GAA AGT GAG 1728
His Thr Tyr Leu Gly Asn Pro Glu Asn Glu Ser Glu
555 560

ATA TAT CCA GAT TTC GTG ACG ATC GCC AAA GGG TTC 1764
Ile Tyr Pro Asp Phe Val Thr Ile Ala Lys Gly Phe
565 570 575

AAC ATT CCA GCG GTC CGT GTG ACA AAG AAG AAC GAA 1800
Asn Ile Pro Ala Val Arg Val Thr Lys Lys Asn Glu
580 585

EP 0 525 384 A2

| | | |
|----|---|------|
| | GTC CGC GCA GCG ATA AAG AAG ATG CTC GAG ACT CCA | 1836 |
| | Val Arg Ala Ala Ile Lys Lys Met Leu Glu Thr Pro | |
| 5 | 590 595 600 | |
| | GGG CCG TAC CTC TTG GAT ATA ATC GTC CCA CAC CAG | 1872 |
| | Gly Pro Tyr Leu Leu Asp Ile Ile Val Pro His Gln | |
| 10 | 605 610 | |
| | GAG CAT GTG TTG CCT ATG ATC CCT AAT GGT GGG GCT | 1908 |
| 15 | Glu His Val Leu Pro Met Ile Pro Asn Gly Gly Ala | |
| | 615 620 | |
| | TTC AAG GAT ATG ATC CTG GAT GGT GAT GGC AGG ACT | 1944 |
| 20 | Phe Lys Asp Met Ile Leu Asp Gly Asp Gly Arg Thr | |
| | 625 630 635 | |
| 25 | GTG TAC | 1950 |
| | Val Tyr | |
| | 638 | |
| 30 | | |
| | TGATCTAAAA TCCAGCAAG | 1969 |
| 35 | | |
| 40 | | |
| 45 | | |
| 50 | | |
| 55 | | |

5

10

15

Original Source Organism: Zea mays

20

25

30

35

40

49

S

ATG GCT CCC CCG GCC ACC CCG CTC CGG CCG TGG GGC 216
Met Ala Pro Pro Ala Thr Pro Leu Arg Pro Trp Gly
50 55 60

| | | |
|----|---|-----|
| 5 | CCC ACC GAT CCC CGC AAG GGC GCC GAC ATC CTC GTC | 252 |
| | Pro Thr Asp Pro Arg Lys Gly Ala Asp Ile Leu Val | |
| | 65 70 | |
| 10 | GAG TCC CTC GAG CGC TGC GGC GTC CGC GAC GTC TTC | 288 |
| | Glu Ser Leu Glu Arg Cys Gly Val Arg Asp Val Phe | |
| | 75 80 | |
| 15 | GCC TAC CCC GGC GGC GCG TCC ATG GAG ATC CAC CAG | 324 |
| | Ala Tyr Pro Gly Gly Ala Ser Met Glu Ile His Gln | |
| | 85 90 95 | |
| 20 | GCA CTC ACC CGC TCC CCC GTC ATC GCC AAC CAC CTC | 360 |
| | Ala Leu Thr Arg Ser Pro Val Ile Ala Asn His Leu | |
| | 100 105 | |
| 25 | TTC CGC CAC GAG CAA GGG GAG GCC TTT GCG GCC TCC | 396 |
| | Phe Arg His Glu Gln Gly Glu Ala Phe Ala Ala Ser | |
| 30 | 110 115 120 | |
| 35 | GGC TAC GCG CGC TCC TCG GGC CGC GTC GGC GTC TGC | 432 |
| | Gly Tyr Ala Arg Ser Ser Gly Arg Val Gly Val Cys | |
| | 125 130 | |
| 40 | ATC GCC ACC TCC GGC CCC GGC GCC ACC AAC CTT GTC | 468 |
| | Ile Ala Thr Ser Gly Pro Gly Ala Thr Asn Leu Val | |
| | 135 140 | |
| 45 | TCC GCG CTC GCC GAC GCG CTG CTC GAT TCC GTC CCC | 504 |
| | Ser Ala Leu Ala Asp Ala Leu Asp Ser Val Pro | |
| | 145 150 155 | |
| 50 | ATG GTC GCC ATC ACG GGA CAG GTG CCG CGA CGC ATG | 540 |
| | Met Val Ala Ile Thr Gly Gln Val Pro Arg Arg Met | |
| 55 | 160 165 | |

| | | |
|----|---|-----|
| 5 | ATT GGC ACC GAC GCC TTC CAG GAG ACG CCC ATC GTC | 576 |
| | Ile Gly Thr Asp Ala Phe Gln Glu Thr Pro Ile Val | |
| | 170 175 180 | |
| 10 | GAG GTC ACC CGC TCC ATC ACC AAG CAC AAC TAC CTG | 612 |
| | Glu Val Thr Arg Ser Ile Thr Lys His Asn Tyr Leu | |
| | 185 190 | |
| 15 | GTC CTC GAC GTC GAC GAC ATC CCC CGC GTC GTG CAG | 648 |
| | Val Leu Asp Val Asp Asp Ile Pro Arg Val Val Gln | |
| | 195 200 | |
| 20 | GAG GCT TTC TTC CTC GCC TCC TCT GGT CGA CCG GGG | 684 |
| | Glu Ala Phe Phe Leu Ala Ser Ser Gly Arg Pro Gly | |
| | 205 210 215 | |
| 25 | CCG GTG CTT GTC GAC ATC CCC AAG GAC ATC CAG CAG | 720 |
| | Pro Val Leu Val Asp Ile Pro Lys Asp Ile Gln Gln | |
| | 220 225 | |
| 30 | CAG ATG GCG GTG CCT GTC TGG GAC AAG CCC ATG AGT | 756 |
| | Gln Met Ala Val Pro Val Trp Asp Lys Pro Met Ser | |
| | 230 235 240 | |
| 35 | CTG CCT GGG TAC ATT GCG CGC CTT CCC AAG CCC CCT | 792 |
| | Leu Pro Gly Tyr Ile Ala Arg Leu Pro Lys Pro Pro | |
| | 245 250 | |
| 40 | GCG ACT GAG TTG CTT GAG CAG GTG CTG CGT CTT GTT | 828 |
| | Ala Thr Glu Leu Leu Glu Gln Val Leu Arg Leu Val | |
| | 255 260 | |
| 45 | GGT GAA TCC CGG CGC CCT GTT CTT TAT GTT GGC GGT | 864 |
| | Gly Glu Ser Arg Arg Pro Val Leu Tyr Val Gly Gly | |
| | 265 270 275 | |
| 50 | | |
| 55 | | |

| | | |
|----|--|------|
| 5 | GCG TGC GCA GCA TCT GGT GAG GAG TTG CGA CGC TTT Ala Cys Ala Ala Ser Gly Glu Glu Leu Arg Arg Phe | 900 |
| | 280 285 | |
| 10 | GTG GAG CTG ACT GGA ATC CCG GTC ACA ACT ACT CTT Val Glu Leu Thr Gly Ile Pro Val Thr Thr Thr Leu | 936 |
| | 290 295 300 | |
| 15 | ATG GGC CTC GGC AAC TTC CCC AGC GAC GAC CCA CTG Met Gly Leu Gly Asn Phe Pro Ser Asp Asp Pro Leu | 972 |
| | 305 310 | |
| 20 | TCT CTG CGC ATG CTA GGT ATG CAT GGC ACG GTG TAT Ser Leu Arg Met Leu Gly Met His Gly Thr Val Tyr | 1008 |
| | 315 320 | |
| 25 | | |
| 30 | GCA AAT TAT GCA GTG GAT AAG GCC GAT CTG TTG CTT Ala Asn Tyr Ala Val Asp Lys Ala Asp Leu Leu Leu | 1044 |
| | 325 330 335 | |
| 35 | GCA CTT GGT GTG CGG TTT GAT GAT CGT GTG ACA GGG Ala Leu Gly Val Arg Phe Asp Asp Arg Val Thr Gly | 1080 |
| | 340 345 | |
| 40 | AAG ATT GAG GCT TTT GCA AGC AGG GCT AAG ATT GTG Lys Ile Glu Ala Phe Ala Ser Arg Ala Lys Ile Val | 1116 |
| | 350 355 360 | |
| 45 | CAC GTT GAT ATT GAT CCG GCT GAG ATT GGC AAG AAC His Val Asp Ile Asp Pro Ala Glu Ile Gly Lys Asn | 1152 |
| | 365 370 | |
| 50 | AAG CAG CCA CAT GTG TCC ATC TGT GCA GAT GTT AAG Lys Gln Pro His Val Ser Ile Cys Ala Asp Val Lys | 1188 |
| 55 | 375 380 | |

CTT GCT TTG CAG GGC ATG AAT GCT CTT CTT GAA GGA 1224
Leu Ala Leu Gln Gly Met Asn Ala Leu Leu Glu Gly
385 390 395

AGC ACA TCA AAG AAG AGC TTT GAC TTT GGC TCA TGG 1260
Ser Thr Ser Lys Lys Ser Phe Asp Phe Gly Ser Trp
400 405

AAC GAT GAG TTG GAT CAG CAG AAG AGG GAA TTC CCC 1296
Asn Asp Glu Leu Asp Gln Gln Lys Arg Glu Phe Pro
410 415 420

CTT GGG TAT AAA ACA TCT AAT GAG GAG ATC CAG CCA 1332
Leu Gly Tyr Lys Thr Ser Asn Glu Glu Ile Gln Pro
425 430

CAA TAT GCT ATT CAG GTT CTT GAT GAG CTG ACG AAA 1368
Gln Tyr Ala Ile Gln Val Leu Asp Glu Leu Thr Lys
435 440

GGC GAG GCC ATC ATC GGC ACA GGT GTT GGG CAG CAC 1404
Gly Glu Ala Ile Ile Gly Thr Gly Val Gly Gln His
445 450 455

CAT ATG TGG GCG GCA CAG TAC TAC ACT TAC AAG CGG 1440
Gln Met Trp Ala Ala Gln Tyr Tyr Thr Tyr Lys Arg
460 465

CCA AGG CAG TGG TTG TCT TCA GCT GGT CTT GGG GCT 1476
Pro Arg Gln Trp Leu Ser Ser Ala Gly Leu Gly Ala
470 475 480

ATG GGA TTT GGT TTG CCG GCT GCT GCT GGT GCT TCT 1512
Met Gly Phe Gly Leu Pro Ala Ala Ala Gly Ala Ser
 485 490

| | | |
|----|---|------|
| | GTG GCC AAC CCA GGT GTT ACT GTT GTT GAC ATC GAT | 1548 |
| | Val Ala Asn Pro Gly Val Thr Val Val Asp Ile Asp | |
| 5 | 495 500 | |
| | GGA GAT GGT AGC TTT CTC ATG AAC GTT CAG GAG CTA | 1584 |
| | Gly Asp Gly Ser Phe Leu Met Asn Val Gln Glu Leu | |
| 10 | 505 510 515 | |
| | GCT ATG ATC CGA ATT GAG AAC CTC CCG GTG AAG GTC | 1620 |
| 15 | Ala Met Ile Arg Ile Glu Asn Leu Pro Val Lys Val | |
| | 520 525 | |
| | TTT GTG CTA AAC AAC CAG CAC CTG GGG ATG GTG GTG | 1656 |
| 20 | Phe Val Leu Asn Asn Gln His Leu Gly Met Val Val | |
| | 530 535 540 | |
| | CAG TGG GAG GAC AGG TTC TAT AAG GCC AAC AGA GCG | 1692 |
| 25 | Gln Trp Glu Asp Arg Phe Tyr Lys Ala Asn Arg Ala | |
| | 545 550 | |
| 30 | CAC ACA TAC TTG GGA AAC CCA GAG AAT GAA AGT GAG | 1728 |
| | His Thr Tyr Leu Gly Asn Pro Glu Asn Glu Ser Glu | |
| 35 | 555 560 | |
| | ATA TAT CCA GAT TTC GTG ACG ATC GCC AAA GGG TTC | 1764 |
| | Ile Tyr Pro Asp Phe Val Thr Ile Ala Lys Gly Phe | |
| 40 | 565 570 575 | |
| | AAC ATT CCA GCG GTC CGT GTG ACA AAG AAG AAC GAA | 1800 |
| 45 | Asn Ile Pro Ala Val Arg Val Thr Lys Lys Asn Glu | |
| | 580 585 | |
| | GTC CGC GCA GCG ATA AAG AAG ATG CTC GAG ACT CCA | 1836 |
| 50 | Val Arg Ala Ala Ile Lys Lys Met Leu Glu Thr Pro | |
| | 590 595 600 | |
| 55 | | |

| | | |
|----|---|------|
| | GGG CCG TAC CTC TTG GAT ATA ATC GTC CCA CAC CAG | 1872 |
| | Gly Pro Tyr Leu Leu Asp Ile Ile Val Pro His Gln | |
| 5 | 605 610 | |
| | GAG CAT GTG TTG CCT ATG ATC CCT AGT GGT GGG GCT | 1908 |
| 10 | Glu His Val Leu Pro Met Ile Pro Ser Gly Gly Ala | |
| | 615 620 | |
| | TTC AAG GAT ATG ATC CTG GAT GGT GAT GGC AGG ACT | 1944 |
| 15 | Phe Lys Asp Met Ile Leu Asp Gly Asp Gly Arg Thr | |
| | 625 630 635 | |
| | GTG TAC | 1950 |
| 20 | Val Tyr | |
| | 638 | |
| 25 | TGATCTAAAA TCCAGCAAG | 1969 |
| 30 | | |
| 35 | | |
| 40 | | |
| 45 | | |
| 50 | | |
| 55 | | |

Sequence ID No.: 3

Sequence Type: Nucleotide and Amino Acid

Sequence Length: 1969 BP's and 638 Amino Acids

Strandedness: Single

Topology: Linear

Original Source Organism: Zea mays

Properties: Herbicide Sensitive AHAS Enzyme

AACCCTCGCG CCGCCTCCGA GACAGCCGCC GCAACC 36

ATG GCC ACC GCC GCC GCC GCG TCT ACC GCG CTC ACT 72

Met Ala Thr Ala Ala Ala Ser Thr Ala Leu Thr
1 5 10

GGC GCC ACT ACC GCT GCG CCC AAG GCG AGG CGC CGG 108

Gly Ala Thr Thr Ala Ala Pro Lys Ala Arg Arg Arg
15 20

GCG CAC CTC CTG GCC ACC GCG GCG GCC CTC GCC GCG 144

Ala His Leu Leu Ala Thr Arg Arg Ala Leu Ala Ala
25 30 35

CCC ATC AGG TGC TCA GCG GCG TCA CCC GCC ATG CCG 180

Pro Ile Arg Cys Ser Ala Ala Ser Pro Ala Met Pro
40 45

ATG GCT CCC CCG GCC ACC CCG CTC CGG CCG TGG GGC 216

Met Ala Pro Pro Ala Thr Pro Leu Arg Pro Trp Gly
50 55 60

| | | |
|----|--|-----|
| 5 | CCC ACC GAG CCC CGC AAG GGT GCT GAC ATC CTC GTC Pro Thr Glu Pro Arg Lys Gly Ala Asp Ile Leu Val | 252 |
| | 65 70 | |
| 10 | GAG TCC CTC GAG CGC TGC GGC GTC CGC GAC GTC TTC Glu Ser Leu Glu Arg Cys Gly Val Arg Asp Val Phe | 288 |
| | 75 80 | |
| 15 | GCC TAC CCC GGC GGC GCG TCC ATG GAG ATC CAC CAG Ala Tyr Pro Gly Gly Ala Ser Met Glu Ile His Gln | 324 |
| | 85 90 95 | |
| 20 | GCA CTC ACC CGC TCC CCC GTC ATC GCC AAC CAC CTC Ala Leu Thr Arg Ser Pro Val Ile Ala Asn His Leu | 360 |
| | 100 105 | |
| 25 | TTC CGC CAC GAG CAA GGG GAG GCC TTT GCC GCC TCC Phe Arg His Glu Gln Gly Glu Ala Phe Ala Ala Ser | 396 |
| 30 | 110 115 120 | |
| 35 | GGC TAC GCG CGC TCC TCG GGC CGC GTC GGC GTC TGC Gly Tyr Ala Arg Ser Ser Gly Arg Val Gly Val Cys | 432 |
| | 125 130 | |
| 40 | ATC GCC ACC TCC GGC CCC GGC GCC ACC AAC CTA GTC Ile Ala Thr Ser Gly Pro Gly Ala Thr Asn Leu Val | 468 |
| | 135 140 | |
| 45 | TCC GCG CTC GCC GAC GCG CTG CTC GAT TCC GTC CCC Ser Ala Leu Ala Asp Ala Leu Leu Asp Ser Val Pro | 504 |
| | 145 150 155 | |
| 50 | ATG GTC GCC ATC ACG GGA CAG GTG CCG CGA CGC ATG Met Val Ala Ile Thr Gly Gln Val Pro Arg Arg Met | 540 |
| 55 | 160 165 | |

| | | |
|----|---|-----|
| | ATT GGC ACC GAC GCC TTC CAG GAG ACG CCC ATC GTC | 576 |
| | Trp Gly Thr Asp Ala Phe Gln Glu Thr Pro Ile Val | |
| 5 | 170 175 180 | |
| | GAG GTC ACC CGC TCC ATC ACC AAG CAC AAC TAC CTG | 612 |
| | Glu Val Thr Arg Ser Ile Thr Lys His Asn Tyr Leu | |
| 10 | 185 190 | |
| | GTC CTC GAC GTC GAC GAC ATC CCC CGC GTC GTG CAG | 648 |
| | Val Leu Asp Val Asp Asp Ile Pro Arg Val Val Gln | |
| 15 | 195 200 | |
| | GAG GCT TTC TTC CTC GCC TCC TCT GGT CGA CCA GGG | 684 |
| | Glu Ala Phe Phe Leu Ala Ser Ser Gly Arg Pro Gly | |
| 20 | 205 210 215 | |
| | CCG GTG CTT GTC GAC ATC CCC AAG GAC ATC CAG CAG | 720 |
| | Pro Val Leu Val Asp Ile Pro Lys Asp Ile Gln Gln | |
| 25 | 220 225 | |
| | CAG ATG GCG GTG CCT GTC TGG GAC AAG CCC ATG AGT | 756 |
| | Gln Met Ala Val Pro Val Trp Asp Lys Pro Met Ser | |
| 30 | 230 235 240 | |
| | CTG CCT GGG TAC ATT GCG CGC CTT CCC AAG CCC CCT | 792 |
| | Leu Pro Gly Tyr Ile Ala Arg Leu Pro Lys Pro Pro | |
| 35 | 245 250 | |
| | GCG ACT GAG TTG CTT GAG CAG GTG CTG CGT CTT GTT | 828 |
| | Ala Thr Glu Leu Leu Glu Gln Val Leu Arg Leu Val | |
| 40 | 255 260 | |
| | GGT GAA TCG CGG CGC CCT GTT CTT TAT GTG GGC GGT | 864 |
| | Gly Glu Ser Arg Arg Pro Val Leu Tyr Val Gly Gly | |
| 45 | 265 270 275 | |
| | | |
| 50 | | |
| 55 | | |

GCG TGC GCA GCA TCT GGT GAG GAG TTG CGA CGC TTT 900
Ala Cys Ala Ala Ser Gly Glu Glu Leu Arg Arg Phe

280 **285**

GTG GAG CTG ACT GGA ATC CCG GTC ACA ACT ACT CTT 936
Val Glu Leu Thr Gly Ile Pro Val Thr Thr Thr Leu

290 295 300

ATG GGC CTC GGC AAC TTC CCC AGC GAC GAC CCA CTG 972
Met Gly Leu Gly Asn Phe Pro Ser Asp Asp Pro Leu

305 310

TCT CTG CGC ATG CTA GGT ATG CAT GGG ACG GTG TAT 1008
Ser Leu Arg Met Leu Gly Met His Gly Thr Val Tyr

315 **320**

GCA AAT TAT GCA GTG GAT AAG GCC GAT CTG TTG CTT 1044
Ala Asn Tyr Ala Val Asp Lys Ala Asp Leu Leu Leu

325

GCA CTT GGT GTG CGG TTT GAT GAT CGT GTG ACA GGG 1080
Ala Leu Gly Val Arg Phe Asp Asp Arg Val Thr Gly

340 345

AAG ATT GAG GCT TTT GCA AGC AGG GCT AAG ATT GTG 1116
Lys Ile Glu Ala Phe Ala Ser Arg Ala Lys Ile Val

350

CAC GTT GAT ATT GAT CCG GCT GAG ATT GGC AAG AAC 1152
His Val Asp Ile Asp Pro Ala Glu Ile Gly Lys Asn

365 **370**

AAG CAG CCA CAT GTG TCC ATC TGT GCA GAT GTT AAG 1188
Lys Gln Pro His Val Ser Ile Cys Ala Asp Val Lys

375 **380**

| | | |
|----|---|------|
| | CTT GCT TTG CAG GGC ATG AAT GCT CTT CTT GAA GGA | 1224 |
| | Leu Ala Leu Gln Gly Met Asn Ala Leu Leu Glu Gly | |
| 5 | 385 390 395 | |
| | AGC ACA TCA AAG AAG AGC TTT GAC TTT GGC TCA TGG | 1260 |
| 10 | Ser Thr Ser Lys Lys Ser Phe Asp Phe Gly Ser Trp | |
| | 400 405 | |
| | AAC GAT GAG TTG GAT CAG CAG AAG AGG GAA TTC CCC | 1296 |
| 15 | Asn Asp Glu Leu Asp Gln Gln Lys Arg Glu Phe Pro | |
| | 410 415 420 | |
| | CTT GGG TAT AAA ACA TCT AAT GAG GAG ATC CAG CCA | 1332 |
| 20 | Leu Gly Tyr Lys Thr Ser Asn Glu Glu Ile Gln Pro | |
| | 425 430 | |
| | CAA TAT GCT ATT CAG GTT CTT GAT GAG CTG ACG AAA | 1368 |
| 25 | Gln Tyr Ala Ile Gln Val Leu Asp Glu Leu Thr Lys | |
| | 435 440 | |
| 30 | GGC GAG GCC ATC ATC GGC ACA GGT GTT GGG CAG CAC | 1404 |
| | Gly Glu Ala Ile Ile Gly Thr Gly Val Gly Gln His | |
| 35 | 445 450 455 | |
| | CAT ATG TGG GCG GCA CAG TAC TAC ACT TAC AAG CGG | 1440 |
| 40 | Gln Met Trp Ala Ala Gln Tyr Tyr Thr Tyr Lys Arg | |
| | 460 465 | |
| | CCA AGG CAG TGG TTG TCT TCA GCT GGT CTT GGG GCT | 1476 |
| 45 | Pro Arg Gln Trp Leu Ser Ser Ala Gly Leu Gly Ala | |
| | 470 475 480 | |
| | ATG GGA TTT GGT TTG CCG GCT GCT GCT GGT GCT TCT | 1512 |
| 50 | Met Gly Phe Gly Leu Pro Ala Ala Ala Gly Ala Ser | |
| | 485 490 | |
| 55 | | |

| | | |
|----|---|------|
| 5 | GTG GCC AAC CCA GGT GTC ACT GTT GTT GAC ATC GAT | 1548 |
| | Val Ala Asn Pro Gly Val Thr Val Val Asp Ile Asp | |
| | 495 500 | |
| 10 | GGA GAT GGT AGC TTT CTC ATG AAC GTT CAG GAG CTA | 1584 |
| | Gly Asp Gly Ser Phe Leu Met Asn Val Gln Glu Leu | |
| | 505 510 515 | |
| 15 | GCT ATG ATC CGA ATT GAG AAC CTC CCA GTG AAG GTC | 1620 |
| | Ala Met Ile Arg Ile Glu Asn Leu Pro Val Lys Val | |
| | 520 525 | |
| 20 | TTT GTG CTA AAC AAC CAG CAC CTG GGG ATG GTG GTG | 1656 |
| | Phe Val Leu Asn Asn Gln His Leu Gly Met Val Val | |
| 25 | 530 535 540 | |
| 30 | CAG TGG GAG GAC AGG TTC TAT AAG GCC AAC AGA GCG | 1692 |
| | Gln Trp Glu Asp Arg Phe Tyr Lys Ala Asn Arg Ala | |
| | 545 550 | |
| 35 | CAC ACA TAC TTG GGA AAC CCA GAG AAT GAA AGT GAG | 1728 |
| | His Thr Tyr Leu Gly Asn Pro Glu Asn Glu Ser Glu | |
| | 555 560 | |
| 40 | ATA TAT CCA GAT TTC GTG ACG ATC GCC AAA GGG TTC | 1764 |
| | Ile Tyr Pro Asp Phe Val Thr Ile Ala Lys Gly Phe | |
| | 565 570 575 | |
| 45 | AAC ATT CCA GCG GTC CGT GTG ACA AAG AAG AAC GAA | 1800 |
| | Asn Ile Pro Ala Val Arg Val Thr Lys Lys Asn Glu | |
| | 580 585 | |
| 50 | GTG CGC GCA GCG ATA AAG AAG ATG CTC GAG ACT CCA | 1836 |
| | Val Arg Ala Ala Ile Lys Lys Met Leu Glu Thr Pro | |
| 55 | 590 595 600 | |

5 GGG CCG TAC CTC TTG GAT ATA ATC GTC CCA CAC CAG 1872
 Gly Pro Tyr Leu Leu Asp Ile Ile Val Pro His Gln
 605 610

10 GAG CAT GTG TTG CCT ATG ATC CCT AGT GGT GGG GCT 1908
 Glu His Val Leu Pro Met Ile Pro Ser Gly Gly Ala
 615 620

15 TTC AAG GAT ATG ATC CTG GAT GGT GAT GGC AGG ACT 1944
 Phe Lys Asp Met Ile Leu Asp Gly Asp Gly Arg Thr
 625 630 635

20 GTG TAC 1950
 Val Tyr
 638

25 TGATCTAAAA TCCAGCAAG 1969

30

Claims

- 35 1. A monocot nucleic acid sequence encoding a functional AHAS enzyme, which enzyme has an amino acid substitution relative to a wild-type monocot AHAS enzyme, and which substitution confers imidazolinone-specific resistance to the enzyme.
- 40 2. The sequence of Claim 1 in which the monocot is corn and the substitution is at position 621 in the wild-type corn AHAS enzyme.
- 45 3. The sequence of Claim 2 in which the substituted amino acid is asparagine.
4. A functional monocot AHAS enzyme which has an amino acid substitution relative to a monocot wild-type AHAS enzyme, and which substitution confers imidazolinone-specific resistance to the enzyme.
5. The enzyme of Claim 4 in which the monocot is corn and the substitution is at position 621 in the wild-type corn AHAS enzyme.
6. The enzyme of Claim 5 in which the substituted amino acid is asparagine.
- 50 7. A transformation vector comprising the nucleic acid of Claim 1.
8. A host cell comprising the nucleic acid sequence of Claim 1, or the vector of Claim 7.
- 55 9. The host cell of Claim 8 which is a plant cell or a bacterial cell.
10. An imidazolinone-specific resistant mature plant containing the nucleic acid sequence of Claim 1, or seed or pollen therefrom.

11. A method of conferring imidazolinone-specific resistance to a plant cell which comprises providing the plant cell with the nucleic acid sequence of Claim 1.
12. A method for growing imidazolinone-specific resistant plants which comprises cultivating a plant which produces the enzyme of Claim 4 in the presence of an inhibitory amount of imidazolinone.
13. A method of selecting host cells successfully transformed with a gene of interest which comprises providing to prospective host cells the gene of interest linked to the nucleic acid sequence of Claim 1, or unlinked but in the presence of the nucleic acid sequence of Claim 1, growing the cells in the presence of an inhibitory amount of imidazolinone and identifying surviving cells as containing the gene of interest.
14. A nucleic acid construct comprising the sequence of Claim 1 linked to a gene encoding an agronomically useful trait.

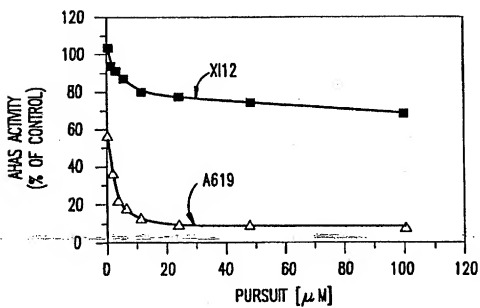


FIG.1A

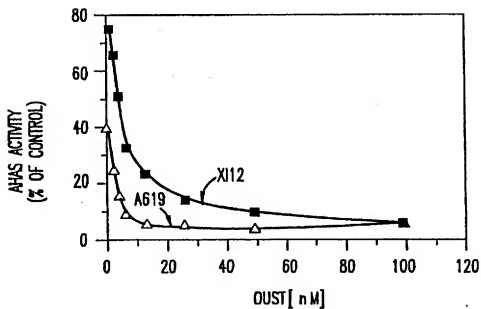


FIG.1B

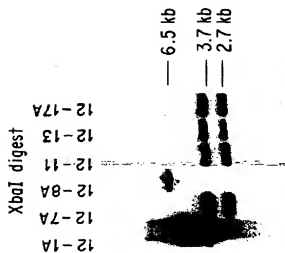


FIG. 2B

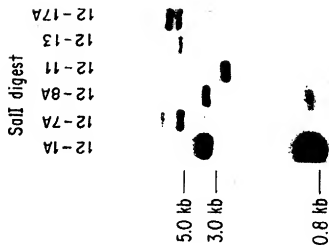


FIG. 2A

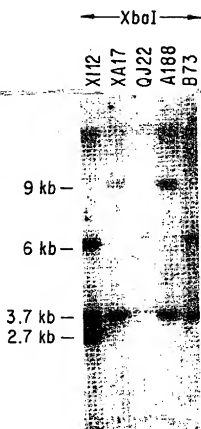


FIG. 3

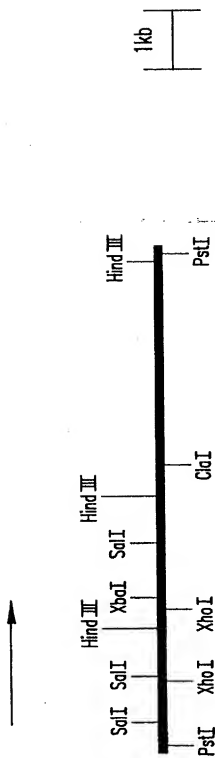


FIG.4

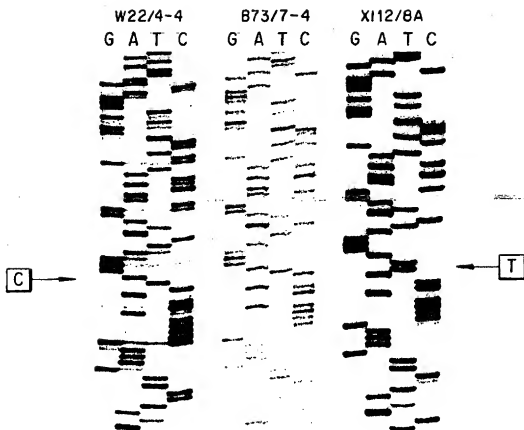


FIG. 5A

W22/1A and B73/7-4
sequence: 5'TAGTG3'
3'ATCTG5'

XI12/8A sequence: 5'TAATG3'
3'ATTAC5'

FIG. 5B

| | | | | | |
|---|-----|-----|-----|-----|-----|
| 40 | 50 | 60 | 70 | 80 | 90 |
| × | × | × | × | × | × |
| ATG GCC ACC GCC GCC GCC GCG TCT ACC GCG CTC ACT GGC GCC ACT ACC GCT GCG | | | | | |
| Met Ala Thr Ala Ala Ala Ala Ser Thr Ala Leu Thr Gly Ala Thr Thr Ala Ala | | | | | |
| 100 | 110 | 120 | 130 | 140 | |
| × | × | × | × | × | |
| CCC AAG GCG AGG CCG GCG GCG CAC CTC CTG GCC ACC GCG CCG GCC CTC GCC GCG | | | | | |
| Pro Lys Ala Arg Arg Arg Ala His Leu Leu Ala Thr Arg Arg Ala Leu Ala Ala | | | | | |
| 150 | 160 | 170 | 180 | 190 | |
| × | × | × | × | × | |
| CCC ATC AGG TGC TCA GCG GCG TCA CCC GCC ATG CCG ATG GCT CCC CCG GCC ACC | | | | | |
| Pro Ile Arg Cys Ser Ala Ala Ser Pro Ala Met Pro Met Ala Pro Pro Ala Thr | | | | | |
| 200 | 210 | 220 | 230 | 240 | 250 |
| × | × | × | × | × | × |
| CCG CTC CCG CCG TGG GGC CCC ACC GAT CCC CCG AAG GGC GCC GAC ATC CTC GTC | | | | | |
| Pro Leu Arg Pro Trp Gly Pro Thr Asp Pro Arg Lys Gly Ala Asp Ile Leu Val | | | | | |
| 260 | 270 | 280 | 290 | 300 | |
| × | × | × | × | × | |
| GAG TCC CTC GAG CCG TGC GGC GTC CCG GAC GTC TTC GCC TAC CCC GGC GGC GCG | | | | | |
| Glu Ser Leu Glu Arg Cys Gly Val Arg Asp Val Phe Ala Tyr Pro Gly Gly Ala | | | | | |
| 310 | 320 | 330 | 340 | 350 | 360 |
| × | × | × | × | × | × |
| TCC ATG GAG ATC CAC CAG GCA CTC ACC CCG TCC CCC GTC ATC GCC AAC CAC CTC | | | | | |
| Ser Met Glu Ile His Gln Ala Leu Thr Arg Ser Pro Val Ile Ala Asn His Leu | | | | | |
| 370 | 380 | 390 | 400 | 410 | |
| × | × | × | × | × | |
| TTC CCG CAC GAG CAA GGG GAG GCC TTT GCG GCC TCC GGC TAC GCG CCG TCC TCG | | | | | |
| Phe Arg His Glu Gln Gly Glu Ala Phe Ala Ala Ser Gly Tyr Ala Arg Ser Ser | | | | | |

FIG.6A

| | | | | | |
|---|-----|-----|-----|-----|-----|
| 420 | 430 | 440 | 450 | 460 | |
| x | x | x | x | x | |
| GGC CGC GTC GGC GTC TGC ATC GCC ACC TCC GGC CCC GGC GCC ACC AAC CTT GTC | | | | | |
| Gly Arg Val Gly Val Cys Ile Ala Thr Ser Gly Pro Gly Ala Thr Asn Leu Val | | | | | |
| 470 | 480 | 490 | 500 | 510 | 520 |
| x | x | x | x | x | x |
| TCC GCG CTC GCC GAC GCG CTG CTC GAT TCC GTC CCC ATG GTC GCC ATC ACG GGA | | | | | |
| Ser Ala Leu Ala Asp Ala Leu Leu Asp Ser Val Pro Met Val Ala Ile Thr Gly | | | | | |
| 530 | 540 | 550 | 560 | 570 | |
| x | x | x | x | x | |
| CAG GTG CCG CGA CGC ATG ATT GGC ACC GAC GCC TTC CAG GAG ACG CCC ATC GTC | | | | | |
| Gln Val Pro Arg Arg Met Ile Gly Thr Asp Ala Phe Gln Glu Thr Pro Ile Val | | | | | |
| 580 | 590 | 600 | 610 | 620 | 630 |
| x | x | x | x | x | x |
| GAG GTC ACC CGC TCC ATC ACC AAG CAC AAC TAC CTG GTC CTC GAC GTC GAC GAC | | | | | |
| Glu Val Thr Arg Ser Ile Thr Lys His Asn Tyr Leu Val Leu Asp Val Asp Asp | | | | | |
| 640 | 650 | 660 | 670 | 680 | |
| x | x | x | x | x | |
| ATC CCC CGC GTC GTG CAG GAG GCT TTC TTC CTC GCC TCC TCT GGT CGA CCG GGG | | | | | |
| Ile Pro Arg Val Val Gln Glu Ala Phe Phe Leu Ala Ser Ser Gly Arg Pro Gly | | | | | |
| 690 | 700 | 710 | 720 | 730 | |
| x | x | x | x | x | |
| CCG GTG CTT GTC GAC ATC CCC AAG GAC ATC CAG CAG CAG ATG GCG GTG CCT GTC | | | | | |
| Pro Val Leu Val Asp Ile Pro Lys Asp Ile Gln Gln Gln Met Ala Val Pro Val | | | | | |
| 740 | 750 | 760 | 770 | 780 | 790 |
| x | x | x | x | x | x |
| TGG GAC AAG CCC ATG AGT CTG CCT GGG TAC ATT GCG CGC CTT CCC AAG CCC CCT | | | | | |
| Trp Asp Lys Pro Met Ser Leu Pro Gly Tyr Ile Ala Arg Leu Pro Lys Pro Pro | | | | | |

FIG.6B

| | | | | | |
|---|------|------|------|------|------|
| 800 | 810 | 820 | 830 | 840 | |
| × | × | × | × | × | |
| GGG ACT GAG TTG CTT GAG CAG GTG CTG CGT CTT GTT GGT GAA TCC CGG CGC CCT | | | | | |
| Ala Thr Glu Leu Leu Glu Gln Val Leu Arg Leu Val Gly Glu Ser Arg Arg Pro | | | | | |
| 850 | 860 | 870 | 880 | 890 | 900 |
| × | × | × | × | × | × |
| GTT CTT TAT GTT GGC GGT GCG TGC GCA GCA TCT GGT GAG GAG TTG CGA CGC TTT | | | | | |
| Val Leu Tyr Val Gly Gly Ala Cys Ala Ala Ser Gly Glu Glu Leu Arg Arg Phe | | | | | |
| 910 | 920 | 930 | 940 | 950 | |
| × | × | × | × | × | |
| GTG GAG CTG ACT GGA ATC CCG GTC ACA ACT ACT CTT ATG GGC CTC GGC AAC TTC | | | | | |
| Val Glu Leu Thr Gly Ile Pro Val Thr Thr Thr Leu Met Gly Leu Gly Asn Phe | | | | | |
| 960 | 970 | 980 | 990 | 1000 | |
| × | × | × | × | × | |
| CCC AGC GAC GAC CCA CTG TCT CTG CGC ATG CTA GGT ATG CAT GGC ACG GTG TAT | | | | | |
| Pro Ser Asp Asp Pro Leu Ser Leu Arg Met Leu Gly Met His Gly Thr Val Tyr | | | | | |
| 1010 | 1020 | 1030 | 1040 | 1050 | 1060 |
| × | × | × | × | × | × |
| GCA AAT TAT GCA GTG GAT AAG GCC GAT CTG TTG CTT GCA CTT GGT GTG CGG TTT | | | | | |
| Ala Asn Tyr Ala Val Asp Lys Ala Asp Leu Leu Leu Ala Leu Gly Val Arg Phe | | | | | |
| 1070 | 1080 | 1090 | 1100 | 1110 | |
| × | × | × | × | × | |
| GAT GAT CGT GTG ACA GGG AAG ATT GAG GCT TTT GCA AGC AGG GCT AAG ATT GTG | | | | | |
| Asp Asp Arg Val Thr Gly Lys Ile Glu Ala Phe Ala Ser Arg Ala Lys Ile Val | | | | | |
| 1120 | 1130 | 1140 | 1150 | 1160 | 1170 |
| × | × | × | × | × | × |
| CAC GTT GAT ATT GAT CCG GCT GAG ATT GGC AAG AAC AAG CAG CCA CAT GTG TCC | | | | | |
| His Val Asp Ile Asp Pro Ala Glu Ile Gly Lys Asn Lys Gln Pro His Val Ser | | | | | |

FIG.6C

| | | | | | |
|---|------|------|------|------|------|
| 1180 | 1190 | 1200 | 1210 | 1220 | |
| x | x | x | x | x | |
| ATC TGT GCA GAT GTT AAG CTT GCT TTG CAG GGC ATG AAT GCT CTT CTT GAA GGA | | | | | |
| Ile Cys Ala Asp Val Lys Leu Ala Leu Gln Gly Met Asn Ala Leu Leu Glu Gly | | | | | |
| 1230 | 1240 | 1250 | 1260 | 1270 | |
| x | x | x | x | x | |
| AGC ACA TCA AAG AAG AGC TTT GAC TTT GGC TCA TGG AAC GAT GAG TTG GAT CAG | | | | | |
| Ser Thr Ser Lys Lys Ser Phe Asp Phe Gly Ser Trp Asn Asp Glu Leu Asp Gln | | | | | |
| 1280 | 1290 | 1300 | 1310 | 1320 | 1330 |
| x | x | x | x | x | x |
| CAG AAG AGG GAA TTC CCC CTT GGG TAT AAA ACA TCT AAT GAG GAG ATC CAG CCA | | | | | |
| Gln Lys Arg Glu Phe Pro Leu Gly Tyr Lys Thr Ser Asn Glu Glu Ile Gln Pro | | | | | |
| 1340 | 1350 | 1360 | 1370 | 1380 | |
| x | x | x | x | x | |
| CAA TAT GCT ATT CAG GTT CTT GAT GAG CTG ACG AAA GGC GAG GCC ATC ATC GGC | | | | | |
| Gln Tyr Ala Ile Gln Val Leu Asp Glu Leu Thr Lys Gly Glu Ala Ile Ile Gly | | | | | |
| 1390 | 1400 | 1410 | 1420 | 1430 | 1440 |
| x | x | x | x | x | x |
| ACA GGT GTT GGG CAG CAC CAG ATG TGG GCG GCA CAG TAC TAC ACT TAC AAG CGG | | | | | |
| Thr Gly Val Gly Gln His Gln Met Trp Ala Ala Gln Tyr Tyr Thr Tyr Lys Arg | | | | | |
| 1450 | 1460 | 1470 | 1480 | 1490 | |
| x | x | x | x | x | |
| CCA AGG CAG TGG TTG TCT TCA GCT GGT CTT GGG GCT ATG GGA TTT GGT TTG CCG | | | | | |
| Pro Arg Gln Trp Leu Ser Ser Ala Gly Leu Gly Ala Met Gly Phe Gly Leu Pro | | | | | |
| 1500 | 1510 | 1520 | 1530 | 1540 | |
| x | x | x | x | x | |
| GCT GCT GCT GGT GCT TCT GTG GCC AAC CCA GGT GTT ACT GTT GTT GAC ATC GAT | | | | | |
| Ala Ala Ala Gly Ala Ser Val Ala Asn Pro Gly Val Thr Val Val Asp Ile Asp | | | | | |
| 1550 | 1560 | 1570 | 1580 | 1590 | 1600 |
| x | x | x | x | x | x |
| GGA GAT GGT AGC TTT CTC ATG AAC GTT CAG GAG CTA GCT ATG ATC CGA ATT GAG | | | | | |
| Gly Asp Gly Ser Phe Leu Met Asn Val Gln Glu Leu Ala Met Ile Arg Ile Glu | | | | | |

FIG.6D

| | | | | | |
|---|------|------|------|------|------|
| 1610 | 1620 | 1630 | 1640 | 1650 | |
| × | × | × | × | × | |
| AAC CTC CCG GTG AAG GTC TTT GTG CTA AAC AAC CAG CAC CTG GGG ATG GTG GTG | | | | | |
| Asn Leu Pro Val Lys Val Phe Val Leu Asn Asn Gln His Leu Gly Met Val Val | | | | | |
| 1660 | 1670 | 1680 | 1690 | 1700 | 1710 |
| × | × | × | × | × | × |
| CAG TGG GAG GAC AGG TTC TAT AAG GCC AAC AGA GCG CAC ACA TAC TTG GGA AAC | | | | | |
| Gln Trp Glu Asp Arg Phe Tyr Lys Ala Asn Arg Ala His Thr Tyr Leu Gly Asn | | | | | |
| 1720 | 1730 | 1740 | 1750 | 1760 | |
| × | × | × | × | × | |
| CCA GAG AAT GAA AGT GAG ATA TAT CCA GAT TTC GTG ACG ATC GCC AAA GGG TTC | | | | | |
| Pro Glu Asn Glu Ser Glu Ile Tyr Pro Asp Phe Val Thr Ile Ala Lys Gly Phe | | | | | |
| 1770 | 1780 | 1790 | 1800 | 1810 | |
| × | × | × | × | × | |
| AAC ATT CCA GCG GTC CGT GTG ACA AAG AAG AAC GAA GTC CCG GCA GCG ATA AAG | | | | | |
| Asn Ile Pro Ala Val Arg Val Thr Lys Lys Asn Glu Val Arg Ala Ala Ile Lys | | | | | |
| 1820 | 1830 | 1840 | 1850 | 1860 | 1870 |
| × | × | × | × | × | × |
| AAG ATG CTC GAG ACT CCA GGG CCG TAC CTC TTG GAT ATA ATC GTC CCA CAC CAG | | | | | |
| Lys Met Leu Glu Thr Pro Gly Pro Tyr Leu Leu Asp Ile Ile Val Pro His Gln | | | | | |
| 1880 | 1890 | 1900 | 1910 | 1920 | |
| × | × | × | × | × | |
| GAG CAT GTG TTG CCT ATG ATC CCT AAT GGT GGG GCT TTC AAG GAT ATG ATC CTG | | | | | |
| Glu His Val Leu Pro Met Ile Pro Asn Gly Gly Ala Phe Lys Asp Met Ile Leu | | | | | |
| 1930 | 1940 | 1950 | 1960 | | |
| × | × | × | × | | |
| GAT GGT GAT GGC AGG ACT GTG TAC TGATC TAAAA TCCAG CAAG | | | | | |
| Asp Gly Asp Gly Arg Thr Val Tyr | | | | | |

FIG.6E

| | 10 | 20 | 30 | 40 | 50 | 60 |
|---------|-------|-------|-------|-------|-------|-------|
| X112/8A | AACCC | TCGGC | CCGCC | TCCGA | GACAG | CCGCC |
| | GCAAC | CATGG | CCACC | GCCGC | CGCCG | CGTCT |
| W22/1A | AACCC | TCGGC | CCGCC | TCCGA | GACAG | CCGCC |
| | GCAAC | CATGG | CCACC | GCCGC | CGCCG | CGTCT |
| B73/7-4 | AACCC | TCGGC | CCGCC | TCCGA | GACAG | CCGCC |
| | GCAAC | CATGG | CCACC | GCCGC | CGCCG | CGTCT |
| X112/8A | AACCC | TCGGC | CCGCC | TCCGA | GACAG | CCGCC |
| | GCAAC | CATGG | CCACC | GCCGC | CGCCG | CGTCT |
| | 70 | 80 | 90 | 100 | 110 | 120 |
| X112/8A | ACCGC | GCTCA | CTGGC | GCCAC | TACCG | CTGGC |
| | CCCAA | GCGCA | GCGGC | GCGGC | GCACC | TCCTG |
| W22/1A | ACCGC | GCTCA | CTGGC | GCCAC | TACCG | CTGGC |
| | CCCAA | GCGCA | GCGGC | GCGGC | GCACC | TCCTG |
| B73/7-4 | ACCGC | GCTCA | CTGGC | GCCAC | TACCG | CTGGC |
| | CCCAA | GCGCA | GCGGC | GCGGC | GCACC | TCCTG |
| X112/8A | ACCGC | GCTCA | CTGGC | GCCAC | TACCG | CTGGC |
| | CCCAA | GCGCA | GCGGC | GCGGC | GCACC | TCCTG |
| | 130 | 140 | 150 | 160 | 170 | 180 |
| X112/8A | GCCAC | CCGCC | GCGCC | CTCGC | CGCGC | CCATC |
| | AGGTG | CTCAG | CGGCG | TCACC | CGCCA | TGCCG |
| W22/1A | GCCAC | CCGCC | GCGCC | CTCGC | CGCGC | CCATC |
| | AGGTG | CTCAG | CGGCG | TCACC | CGCCA | TGCCG |
| B73/7-4 | GCCAC | CCGCC | GCGCC | CTCGC | CGCGC | CCATC |
| | AGGTG | CTCAG | CGGCG | TCACC | CGCCA | TGCCG |
| X112/8A | GCCAC | CCGCC | GCGCC | CTCGC | CGCGC | CCATC |
| | AGGTG | CTCAG | CGGCG | TCACC | CGCCA | TGCCG |
| | 190 | 200 | 210 | 220 | 230 | 240 |
| X112/8A | ATGGC | TCCCC | CGGCC | ACCCC | GCTCC | GGCGG |
| | TGGGG | CCCCA | CCGAT | CCCCG | CAAGG | GCGCC |
| W22/1A | ATGGC | TCCCC | CGGCC | ACCCC | GCTCC | GGCGG |
| | TGGGG | CCCCA | CCGAT | CCCCG | CAAGG | GCGCC |
| B73/7-4 | ATGGC | TCCCC | CGGCC | ACCCC | GCTCC | GGCGG |
| | TGGGG | CCCCA | CCGAG | CCCCG | CAAGG | GtGct |
| X112/8A | ATGGC | TCCCC | CGGCC | ACCCC | GCTCC | GGCGG |
| | TGGGG | CCCCA | CCGAT | CCCCG | CAAGG | GCGCC |

FIG.7A

| | 250 | 260 | 270 | 280 | 290 | 300 |
|----------|---|-----|-----|-----|-----|-----|
| X112/8A | GACAT CCTCG TCGAG TCCCT CGAGC GCTGC GGGGT CCGCG ACGTC TTCGC CTACC CCGGC | | | | | |
| W22/1A | GACAT CCTCG TCGAG TCCCT CGAGC GCTGC GGGGT CCGCG ACGTC TTCGC CTACC CCGGC | | | | | |
| B73/7-4 | 11111 11111 11111 11111 11111 11111 11111 11111 11111 11111 11111 11111 | | | | | |
| X112/8A | GACAT CCTCG TCGAG TCCCT CGAGC GCTGC GGGGT CCGCG ACGTC TTCGC CTACC CCGGC | | | | | |
| | | | | | | |
| | 310 | 320 | 330 | 340 | 350 | 360 |
| X112/8A | GGCGC GTCCA TGGAG ATCCA CCAGG CACTC ACCCG CTCCC CCGTC ATCGC CAACC ACCTC | | | | | |
| W22/1A | GGCGC GTCCA TGGAG ATCCA CCAGG CACTC ACCCG CTCCC CCGTC ATCGC CAACC ACCTC | | | | | |
| XB73/7-4 | 11111 11111 11111 11111 11111 11111 11111 11111 11111 11111 11111 11111 | | | | | |
| X112/8A | GGCGC GTCCA TGGAG ATCCA CCAGG CACTC ACCCG CTCCC CCGTC ATCGC CAACC ACCTC | | | | | |
| | | | | | | |
| | 370 | 380 | 390 | 400 | 410 | 420 |
| X112/8A | TTCCG CCACG AGCAA GGGGA GGCGT TTGCG GCCTC CGGCT ACGCG CGCTC CTCGG GCGGC | | | | | |
| W22/1A | TTCCG CCACG AGCAA GGGGA GGCGT TTGCG GCCTC CGGCT ACGCG CGCTC CTCGG GCGGC | | | | | |
| B73/7-4 | 11111 11111 11111 11111 11111 1111 11111 11111 11111 11111 11111 11111 | | | | | |
| X112/8A | TTCCG CCACG AGCAA GGGGA GGCGT TTGCG GCCTC CGGCT ACGCG CGCTC CTCGG GCGGC | | | | | |
| | | | | | | |
| | 430 | 440 | 450 | 460 | 470 | 480 |
| X112/8A | GTCGG CGTCT GCATC GCCAC CTCGG GCCCC GGGCG CACCA ACCTT GTCTC CCGCG TCGGC | | | | | |
| W22/1A | GTCGG CGTCT GCATC GCCAC CTCGG GCCCC GGGCG CACCA ACCTT GTCTC CCGCG TCGGC | | | | | |
| B73/7-4 | 11111 11111 11111 11111 11111 11111 11111 11111 1111 11111 11111 11111 | | | | | |
| X112/8A | GTCGG CGTCT GCATC GCCAC CTCGG GCCCC GGGCG CACCA ACCTT GTCTC CCGCG TCGGC | | | | | |
| | | | | | | |

FIG.7B

| | 490 | 500 | 510 | 520 | 530 | 540 |
|---------|---|-----|-----|-----|-----|-----|
| X112/8A | GACGC GCTGC TCGAT TCCGT CCCC TGGTC GCCAT CACGG GACAG GTGCC GCGAC GCATG | | | | | |
| W22/1A | GACGC GCTGC TCGAT TCCGT CCCC TGGTC GCCAT CACGG GACAG GTGCC GCGAC GCATG | | | | | |
| B73/7-4 | 11111 11111 11111 11111 11111 11111 11111 11111 11111 11111 11111 11111 | | | | | |
| X112/8A | GACGC GCTGC TCGAT TCCGT CCCC TGGTC GCCAT CACGG GACAG GTGCC GCGAC GCATG | | | | | |
| | 550 | 560 | 570 | 580 | 590 | 600 |
| X112/8A | ATTGG CACCG ACGCC TTCCA GGAGA CGCCC ATCGT CGAGG TCACC CGCTC CATCA CCAAG | | | | | |
| W22/1A | ATTGG CACCG ACGCC TTCCA GGAGA CGCCC ATCGT CGAGG TCACC CGCTC CATCA CCAAG | | | | | |
| B73/7-4 | 11111 11111 11111 11111 11111 11111 11111 11111 11111 11111 11111 11111 | | | | | |
| X112/8A | ATTGG CACCG ACGCC TTCCA GGAGA CGCCC ATCGT CGAGG TCACC CGCTC CATCA CCAAG | | | | | |
| | 610 | 620 | 630 | 640 | 650 | 660 |
| X112/8A | CACAA CTACC TGGTC CTCGA CGTCG ACGAC ATCCC CCGCG TCGTG CAGGA GGCCT TCTTC | | | | | |
| W22/1A | CACAA CTACC TGGTC CTCGA CGTCG ACGAC ATCCC CCGCG TCGTG CAGGA GGCCT TCTTC | | | | | |
| B73/7-4 | 11111 11111 11111 11111 11111 11111 11111 11111 11111 11111 11111 11111 | | | | | |
| X112/8A | CACAA CTACC TGGTC CTCGA CGTCG ACGAC ATCCC CCGCG TCGTG CAGGA GGCCT TCTTC | | | | | |
| | 670 | 680 | 690 | 700 | 710 | 720 |
| X112/8A | CTCGC CTCCT CTGGT CGACC GGGGC CGGTG CTTGT CGACA TCCCC AAGGA CATCC AGCAG | | | | | |
| W22/1A | CTCGC CTCCT CTGGT CGACC GGGGC CGGTG CTTGT CGACA TCCCC AAGGA CATCC AGCAG | | | | | |
| B73/7-4 | 11111 11111 11111 11111 11111 11111 11111 11111 11111 11111 11111 11111 | | | | | |
| X112/8A | CTCGC CTCCT CTGGT CGACC GGGGC CGGTG CTTGT CGACA TCCCC AAGGA CATCC AGCAG | | | | | |

FIG.7C

| | 730 | 740 | 750 | 760 | 770 | 780 |
|---------|-------------------|-------------------|-------------------|-------------------|-------------|-------|
| X112/8A | CAGAT GCGCG TGCC | GCTCG GGACA AGCCC | ATGAG TCTGC CTGGG | TACAT TCGCG GCCTT | | |
| W22/1A | CAGAT GCGCG TGCC | GCTCG GGACA AGCCC | ATGAG TCTGC CTGGG | TACAT TCGCG GCCTT | | |
| | | | | | | |
| B73/7-4 | CAGAT GCGCG TGCC | GCTCG GGACA AGCCC | ATGAG TCTGC CTGGG | TACAT TCGCG GCCTT | | |
| | | | | | | |
| X112/8A | CAGAT GCGCG TGCC | GCTCG GGACA AGCCC | ATGAG TCTGC CTGGG | TACAT TCGCG GCCTT | | |
| | | | | | | |
| | 790 | 800 | 810 | 820 | 830 | 840 |
| X112/8A | CCCAA GCGCG CTGCG | ACTGA GTTGC | TTGAG CAGGT GCTGC | GCTCT GTTGG TGAAT | CCCGG | |
| | | | | | | |
| X22/1A | CCCAA GCGCG CTGCG | ACTGA GTTGC | TTGAG CAGGT GCTGC | GCTCT GTTGG TGAAT | CCCGG | |
| | | | | | | |
| B73/7-4 | CCCAA GCGCG CTGCG | ACTGA GTTGC | TTGAG CAGGT GCTGC | GCTCT GTTGG TGAAT | GCGCG | |
| | | | | | | |
| X112/8A | CCCAA GCGCG CTGCG | ACTGA GTTGC | TTGAG CAGGT GCTGC | GCTCT GTTGG TGAAT | CCCGG | |
| | | | | | | |
| | 850 | 860 | 870 | 880 | 890 | 900 |
| X112/8A | GCGCC TGTTT TTTAT | GTTGG CGGTG | CGTGC GCAGC | ATCTG GTGAG | GAGTT GCGAC | GCTTT |
| | | | | | | |
| W22/1A | GCGCC TGTTT TTTAT | GTTGG CGGTG | CGTGC GCAGC | ATCTG GTGAG | GAGTT GCGAC | GCTTT |
| | | | | | | |
| B73/7-4 | GCGCC TGTTT TTTAT | GTTGG CGGTG | CGTGC GCAGC | ATCTG GTGAG | GAGTT GCGAC | GCTTT |
| | | | | | | |
| X112/8A | GCGCC TGTTT TTTAT | GTTGG CGGTG | CGTGC GCAGC | ATCTG GTGAG | GAGTT GCGAC | GCTTT |
| | | | | | | |
| | 910 | 920 | 930 | 940 | 950 | 960 |
| X112/8A | GTGGA GCTGA CTGGA | ATCCC GGTCA | CAACT ACTCT | TATGG GCCTC | GGCAA CTTC | CCAGC |
| | | | | | | |
| W22/1A | GTGGA GCTGA CTGGA | ATCCC GGTCA | CAACT ACTCT | TATGG GCCTC | GGCAA CTTC | CCAGC |
| | | | | | | |
| B73/7-4 | GTGGA GCTGA CTGGA | ATCCC GGTCA | CAACT ACTCT | TATGG GCCTC | GGCAA CTTC | CCAGC |
| | | | | | | |
| X112/8A | GTGGA GCTGA CTGGA | ATCCC GGTCA | CAACT ACTCT | TATGG GCCTC | GGCAA CTTC | CCAGC |

FIG.7D

| | 970 | 980 | 990 | 1000 | 1010 | 1020 |
|---------|---|------|------|------|------|------|
| X112/8A | GACGA CCCAC TGTCT CTGCG CATGC TAGGT ATGCA TGGCA CGGTG TATGC AAATT ATGCA | | | | | |
| W22/1A | GACGA CCCAC TGTCT CTGCG CATGC TAGGT ATGCA TGGCA CGGTG TATGC AAATT ATGCA | | | | | |
| B73/7-4 | GACGA CCCAC TGTCT CTGCG CATGC TAGGT ATGCA TGGCA CGGTG TATGC AAATT ATGCA | | | | | |
| X112/8A | GACGA CCCAC TGTCT CTGCG CATGC TAGGT ATGCA TGGCA CGGTG TATGC AAATT ATGCA | | | | | |
| | 1030 | 1040 | 1050 | 1060 | 1070 | 1080 |
| X112/8A | GTGGA TAAGG CCGAT CTGTT GCTTG CACTT GGTGT GCGGT TTGAT GATCG TGTGA CAGGG | | | | | |
| W22/1A | GTGGA TAAGG CCGAT CTGTT GCTTG CACTT GGTGT GCGGT TTGAT GATCG TGTGA CAGGG | | | | | |
| B73/7-4 | GTGGA TAAGG CCGAT CTGTT GCTTG CACTT GGTGT GCGGT TTGAT GATCG cGTGA CAGGG | | | | | |
| X112/8A | GTGGA TAAGG CCGAT CTGTT GCTTG CACTT GGTGT GCGGT TTGAT GATCG TGTGA CAGGG | | | | | |
| | 1090 | 1100 | 1110 | 1120 | 1130 | 1140 |
| X112/8A | AAGAT TGAGG CTTT GCAAG CAGGG CTAAG ATTGT GCACG TTGAT ATTGA TCCGG CTGAG | | | | | |
| W22/1A | AAGAT TGAGG CTTT GCAAG CAGGG CTAAG ATTGT GCACG TTGAT ATTGA TCCGG CTGAG | | | | | |
| B73/7-4 | AAGAT TGAGG CTTT GCAAG CAGGG CTAAG ATTGT GCACG TTGAT ATTGA TCCGG CTGAG | | | | | |
| X112/8A | AAGAT TGAGG CTTT GCAAG CAGGG CTAAG ATTGT GCACG TTGAT ATTGA TCCGG CTGAG | | | | | |
| | 1150 | 1160 | 1170 | 1180 | 1190 | 1200 |
| X112/8A | ATTGG CAAGA ACAAG CAGCC ACATG TGTCG ATCTG TGCAG ATGTT AAGCT TGCTT TGCAG | | | | | |
| W22/1A | ATTGG CAAGA ACAAG CAGCC ACATG TGTCG ATCTG TGCAG ATGTT AAGCT TGCTT TGCAG | | | | | |
| B73/7-4 | ATTGG CAAGA ACAAG CAGCC ACATG TGTCG ATCTG TGCAG ATGTT AAGCT TGCTT TGCAG | | | | | |
| X112/8A | ATTGG CAAGA ACAAG CAGCC ACATG TGTCG ATCTG TGCAG ATGTT AAGCT TGCTT TGCAG | | | | | |

FIG.7E

| | 1210 | 1220 | 1230 | 1240 | 1250 | 1260 |
|---------|-------------------|-------------|-------------|-------------|-------------|-------|
| X112/8A | GGCAT GAATG CTCTT | CTTGA AGGAA | GCACA TCAAA | GAAGA GCTTT | GACTT TGGCT | CATGG |
| W22/1A | GGCAT GAATG CTCTT | CTTGA AGGAA | GCACA TCAAA | GAAGA GCTTT | GACTT TGGCT | CATGG |
| B73/7-4 | GGCAT GAATG CTCTT | CTTGA AGGAA | GCACA TCAAA | GAAGA GCTTT | GACTT TGGCT | CATGG |
| X112/8A | GGCAT GAATG CTCTT | CTTGA AGGAA | GCACA TCAAA | GAAGA GCTTT | GACTT TGGCT | CATGG |
| | 1270 | 1280 | 1290 | 1300 | 1310 | 1320 |
| X112/8A | AACGA TGAGT TGGAT | CAGCA GAAGA | GGGAA TTCCC | CCTTG GGTAT | AAAAC ATCTA | ATGAG |
| W22/1A | AACGA TGAGT TGGAT | CAGCA GAAGA | GGGAA TTCCC | CCTTG GGTAT | AAAAC ATCTA | ATGAG |
| B73/65 | AACGA TGAGT TGGAT | CAGCA GAAGA | GGGAA TTCCC | CCTTG GGTAT | AAAAC ATCTA | ATGAG |
| X112/8A | AACGA TGAGT TGGAT | CAGCA GAAGA | GGGAA TTCCC | CCTTG GGTAT | AAAAC ATCTA | ATGAG |
| | 1330 | 1340 | 1350 | 1360 | 1370 | 1380 |
| X112/8A | GAGAT CCAGC CACAA | TATGC TATTC | AGGTT CTGGA | TGAGC TGAGC | AAAGG CGAGG | CCATC |
| W22/1A | GAGAT CCAGC CACAA | TATGC TATTC | AGGTT CTGGA | TGAGC TGAGC | AAAGG CGAGG | CCATC |
| B73/7-4 | GAGAT CCAGC CACAA | TATGC TATTC | AGGTT CTGGA | TGAGC TGAGC | AAAGG CGAGG | CCATC |
| X112/8A | GAGAT CCAGC CACAA | TATGC TATTC | AGGTT CTGGA | TGAGC TGAGC | AAAGG CGAGG | CCATC |
| | 1390 | 1400 | 1410 | 1420 | 1430 | 1440 |
| X112/8A | ATCGG CACAG GTGTT | GGGCA GCACC | AGATG TGGGC | GGCAC AGTAC | TACAC TTACA | AGCGG |
| W22/1A | ATCGG CACAG GTGTT | GGGCA GCACC | AGATG TGGGC | GGCAC AGTAC | TACAC TTACA | AGCGG |
| B73/7-4 | ATCGG CACAG GTGTT | GGGCA GCACC | AGATG TGGGC | GGCAC AGTAC | TACAC TTACA | AGCGG |
| X112/8A | ATCGG CACAG GTGTT | GGGCA GCACC | AGATG TGGGC | GGCAC AGTAC | TACAC TTACA | AGCGG |

FIG.7F

| | 1450 | 1460 | 1470 | 1480 | 1490 | 1500 |
|---------|---|-------------------------------|------|------|------|------|
| X112/BA | CCAAG GCAGT GGTGG TCCTC AGCTG GTCTT GGGGC | TATGG GATTI GGTIT GCCGG CTGCT | | | | |
| W22/1A | CCAAG GCAGT GGTGG TCCTC AGCTG GTCTT GGGGC | TATGG GATTI GGTIT GCCGG CTGCT | | | | |
| B73/7-4 | CCAAG GCAGT GGTGG TCCTC AGCTG GTCTT GGGGC | TATGG GATTI GGTIT GCCGG CTGCT | | | | |
| X112/BA | CCAAG GCAGT GGTGG TCCTC AGCTG GTCTT GGGGC | TATGG GATTI GGTIT GCCGG CTGCT | | | | |
| | 1510 | 1520 | 1530 | 1540 | 1550 | 1560 |
| X112/BA | GCTGG TGCTT CTGTG GCCAA CCCAG GTGTT ACTGT TGTGG ACATC | GATGG AGATG GTAGC | | | | |
| W22/1A | GCTGG TGCTT CTGTG GCCAA CCCAG GTGTT ACTGT TGTGG ACATC | GATGG AGATG GTAGC | | | | |
| B73/7-4 | GCTGG TGCTT CTGTG GCCAA CCCAG GTGTT ACTGT TGTGG ACATC | GATGG AGATG GTAGC | | | | |
| X112/BA | GCTGG TGCTT CTGTG GCCAA CCCAG GTGTT ACTGT TGTGG ACATC | GATGG AGATG GTAGC | | | | |
| | 1570 | 1580 | 1590 | 1600 | 1610 | 1620 |
| X112/BA | TTTCT CATGA ACGTT CAGGA GCTAG CTATG ATCCG AATTG AGAAC | CTCCC GGTGA AGGTC | | | | |
| W22/1A | TTTCT CATGA ACGTT CAGGA GCTAG CTATG ATCCG AATTG AGAAC | CTCCC GGTGA AGGTC | | | | |
| B73/7-4 | TTTCT CATGA ACGTT CAGGA GCTAG CTATG ATCCG AATTG AGAAC | CTCCC GGTGA AGGTC | | | | |
| X112/BA | TTTCT CATGA ACGTT CAGGA GCTAG CTATG ATCCG AATTG AGAAC | CTCCC GGTGA AGGTC | | | | |
| | 1630 | 1640 | 1650 | 1660 | 1670 | 1680 |
| X112/BA | TTTGT GCTAA ACAAC CAGCA CCTGG GGATG GTGGT GCAGT GGGAG | GACAG GTTCT ATAAG | | | | |
| W22/1A | TTTGT GCTAA ACAAC CAGCA CCTGG GGATG GTGGT GCAGT GGGAG | GACAG GTTCT ATAAG | | | | |
| B73/7-4 | TTTGT GCTAA ACAAC CAGCA CCTGG GGATG GTGGT GCAGT GGGAG | GACAG GTTCT ATAAG | | | | |
| X112/BA | TTTGT GCTAA ACAAC CAGCA CCTGG GGATG GTGGT GCAGT GGGAG | GACAG GTTCT ATAAG | | | | |

FIG.7G

| | 1690 | 1700 | 1710 | 1720 | 1730 | 1740 |
|---------|-------------------|-------------|-------------|-------------|-------------|-------|
| X112/8A | GCCAA CAGAG CGCAC | ACATA CTTGG | GAAAC CCAGA | GAATG AAAGT | GAGAT ATATC | CAGAT |
| W22/1A | GCCAA CAGAG CGCAC | ACATA CTTGG | GAAAC CCAGA | GAATG AAAGT | GAGAT ATATC | CAGAT |
| B73/7-4 | GCCAA CAGAG CGCAC | ACATA CTTGG | GAAAC CCAGA | GAATG AAAGT | GAGAT ATATC | CAGAT |
| X112/8A | GCCAA CAGAG CGCAC | ACATA CTTGG | GAAAC CCAGA | GAATG AAAGT | GAGAT ATATC | CAGAT |
| | 1750 | 1760 | 1770 | 1780 | 1790 | 1800 |
| X112/8A | TTCGT GACGA TCGCC | AAAGG GTTCA | ACATT CCAGC | GGTCC GTGTG | ACAAA GAAGA | ACGAA |
| W22/1A | TTCGT GACGA TCGCC | AAAGG GTTCA | ACATT CCAGC | GGTCC GTGTG | ACAAA GAAGA | ACGAA |
| B73/7-4 | TTCGT GACGA TCGCC | AAAGG GTTCA | ACATT CCAGC | GGTCC GTGTG | ACAAA GAAGA | ACGAA |
| X112/8A | TTCGT GACGA TCGCC | AAAGG GTTCA | ACATT CCAGC | GGTCC GTGTG | ACAAA GAAGA | ACGAA |
| | 1810 | 1820 | 1830 | 1840 | 1850 | 1860 |
| X112/8A | GTCCG CGCAG CGATA | AAGAA GATGC | TCGAG ACTCC | AGGCG CGTAC | CTCTT GGATA | TAATC |
| W22/1A | GTCCG CGCAG CGATA | AAGAA GATGC | TCGAG ACTCC | AGGCG CGTAC | CTCTT GGATA | TAATC |
| B73/7-4 | GTCCG CGCAG CGATA | AAGAA GATGC | TCGAG ACTCC | AGGCG CGTAC | CTCTT GGATA | TAATC |
| X112/8A | GTCCG CGCAG CGATA | AAGAA GATGC | TCGAG ACTCC | AGGCG CGTAC | CTCTT GGATA | TAATC |
| | 1870 | 1880 | 1890 | 1900 | 1910 | 1920 |
| X112/8A | GTCCC ACACC AGGAG | CATGT GTTGC | CTATG ATCCC | TAATG GTGGG | GCTTT CAAGG | ATATG |
| W22/1A | GTCCC ACACC AGGAG | CATGT GTTGC | CTATG ATCCC | TAATG GTGGG | GCTTT CAAGG | ATATG |
| B73/7-4 | GTCCC ACACC AGGAG | CATGT GTTGC | CTATG ATCCC | TAATG GTGGG | GCTTT CAAGG | ATATG |
| X112/8A | GTCCC ACACC AGGAG | CATGT GTTGC | CTATG ATCCC | TAATG GTGGG | GCTTT CAAGG | ATATG |

FIG.7H

| | 1930 | 1940 | 1950 | 1960 | | | | | | |
|---------|-------|-------|-------|-------|-------|-------|-------|-------|-------|-------|
| XI12/8A | ATCCT | GGATG | GTGAT | GGCAG | GACTG | TGTAC | TGATC | TAAAA | TCCAG | CAAG |
| W22/1A | ATCCT | GGATG | GTGAT | GGCAG | GACTG | TGTAC | TGATC | TAAAA | TCCAG | CAAG> |
| | 11111 | 11111 | 11111 | 11111 | 11111 | 11111 | 11111 | 11111 | 11111 | 11111 |
| B73/7-4 | ATCCT | GGATG | GTGAT | GGCAG | GACTG | TGTAC | TGATC | TAAAA | TCCAG | CAAG> |
| | 11111 | 11111 | 11111 | 11111 | 11111 | 11111 | 11111 | 11111 | 11111 | 11111 |
| XI12/8A | ATCCT | GGATG | GTGAT | GGCAG | GACTG | TGTAC | TGATC | TAAAA | TCCAG | CAAG |

FIG.7 I

| | 10 | 20 | 30 | 40 | 50 | 60 |
|---------|-------|-------|-------|-------|-------|---|
| X112/8A | MATAA | AASTA | LTGAT | TAAPK | ARRRA | HLLAT RRALA APIRC SAASP AMPNA PPATP LRPVG |
| W22/1A | MATAA | AASTA | LTGAT | TAAPK | ARRRA | HLLAT RRALA APIRC SAASP AMPNA PPATP LRPVG |
| B73/7-4 | MATAA | AASTA | LTGAT | TAAPK | ARRRA | HLLAT RRALA APIRC SAASP AMPNA PPATP LRPVG |
| X112/8A | MATAA | AASTA | LTGAT | TAAPK | ARRRA | HLLAT RRALA APIRC SAASP AMPNA PPATP LRPVG |

| | 70 | 80 | 90 | 100 | 110 | 120 |
|---------|-------|-------|-------|-------|-------|---|
| X112/8A | PTDPR | KGADI | LVESL | ERCGV | RDVFA | YPGGA SMEIH QALTR SPVIA NHLFR HEGGE AFAAS |
| W22/1A | PTDPR | KGADI | LVESL | ERCGV | RDVFA | YPGGA SMEIH QALTR SPVIA NHLFR HEGGE AFAAS |
| B73/7-4 | PTDPR | KGADI | LVESL | ERCGV | RDVFA | YPGGA SMEIH QALTR SPVIA NHLFR HEGGE AFAAS |
| X112/8A | PTDPR | KGADI | LVESL | ERCGV | RDVFA | YPGGA SMEIH QALTR SPVIA NHLFR HEGGE AFAAS |

| | 130 | 140 | 150 | 160 | 170 | 180 |
|---------|-------|-------|-------|-------|-------|---|
| X112/8A | GYARS | SGRVG | YCIAT | SGPGA | TNLVS | ALADA LLDVS PMVAI TGOVP RRMIG TDAFQ ETPIV |
| W22/1A | GYARS | SGRVG | YCIAT | SGPGA | TNLVS | ALADA LLDVS PMVAI TGOVP RRMIG TDAFQ ETPIV |
| B73/7-4 | GYARS | SGRVG | YCIAT | SGPGA | TNLVS | ALADA LLDVS PMVAI TGOVP RRMIG TDAFQ ETPIV |
| X112/8A | GYARS | SGRVG | YCIAT | SGPGA | TNLVS | ALADA LLDVS PMVAI TGOVP RRMIG TDAFQ ETPIV |

| | 190 | 200 | 210 | 220 | 230 | 240 |
|---------|-------|-------|-------|-------|-------|---|
| X112/8A | EVTRS | ITKHN | YLVLD | VDDIP | RVVQE | AFFLA SSGRP GPVLV DIPKD IQQQM AYPVV DKPHS |
| W22/1A | EVTRS | ITKHN | YLVLD | VDDIP | RVVQE | AFFLA SSGRP GPVLV DIPKD IQQQM AYPVV DKPHS |
| B73/7-4 | EVTRS | ITKHN | YLVLD | VDDIP | RVVQE | AFFLA SSGRP GPVLV DIPKD IQQQM AYPVV DKPHS |
| X112/8A | EVTRS | ITKHN | YLVLD | VDDIP | RVVQE | AFFLA SSGRP GPVLV DIPKD IQQQM AYPVV DKPHS |

FIG.8A

| | 250 | 260 | 270 | 280 | 290 | 300 |
|---------|-------|-------|-------|-------|-------|---|
| X112/8A | LPGYI | ARLPK | PPATE | LLEQV | LRLVG | ESRRP VLYVG GGCAA SGEEL RRFVE LTGIP VTITL |
| V22/1A | LPGYI | ARLPK | PPATE | LLEQV | LRLVG | ESRRP VLYVG GGCAA SGEEL RRFVE LTGIP VTITL |
| B73/7-4 | LPGYI | ARLPK | PPATE | LLEQV | LRLVG | ESRRP VLYVG GGCAA SGEEL RRFVE LTGIP VTITL |
| X112/8A | LPGYI | ARLPK | PPATE | LLEQV | LRLVG | ESRRP VLYVG GGCAA SGEEL RRFVE LTGIP VTITL |
| | 310 | 320 | 330 | 340 | 350 | 360 |
| X112/8A | MGLGN | FPSDD | PLSLR | MGLMH | GTYYA | NYAVD KADLL LALGV RFDDR VTGKI EAFAS RAKIV |
| V22/1A | MGLGN | FPSDD | PLSLR | MGLMH | GTYYA | NYAVD KADLL LALGV RFDDR VTGKI EAFAS RAKIV |
| B73/7-4 | MGLGN | FPSDD | PLSLR | MGLMH | GTYYA | NYAVD KADLL LALGV RFDDR VTGKI EAFAS RAKIV |
| X112/8A | MGLGN | FPSDD | PLSLR | MGLMH | GTYYA | NYAVD KADLL LALGV RFDDR VTGKI EAFAS RAKIV |
| | 370 | 380 | 390 | 400 | 410 | 420 |
| X112/8A | HVDID | PAEIG | KNKOP | HVSIC | ADVKL | ALQGM NALLE GSTSK KSFDF GSWND ELDDQ KREFP |
| V22/1A | HVDID | PAEIG | KNKOP | HVSIC | ADVKL | ALQGM NALLE GSTSK KSFDF GSWND ELDDQ KREFP |
| B73/7-4 | HVDID | PAEIG | KNKOP | HVSIC | ADVKL | ALQGM NALLE GSTSK KSFDF GSWND ELDDQ KREFP |
| X112/8A | HVDID | PAEIG | KNKOP | HVSIC | ADVKL | ALQGM NALLE GSTSK KSFDF GSWND ELDDQ KREFP |
| | 430 | 440 | 450 | 460 | 470 | 480 |
| X112/8A | LGYKT | SNEEI | QPOYA | IQVLD | ELTKG | EATIG TGVGO HQMVA AQYTT YKRPR QVLSS AGLGA |
| V22/1A | LGYKT | SNEEI | QPOYA | IQVLD | ELTKG | EATIG TGVGO HQMVA AQYTT YKRPR QVLSS AGLGA |
| B73/7-4 | LGYKT | SNEEI | QPOYA | IQVLD | ELTKG | EATIG TGVGO HQMVA AQYTT YKRPR QVLSS AGLGA |
| X112/8A | LGYKT | SNEEI | QPOYA | IQVLD | ELTKG | EATIG TGVGO HQMVA AQYTT YKRPR QVLSS AGLGA |

FIG.8B

| | | | | | | |
|---------|---|-----|-----|-----|-----|-----|
| | 490 | 500 | 510 | 520 | 530 | 540 |
| XI12/8A | MGFL PAAAG ASYAN PGVTY VD1DG DGSFL MNVQE LAMIR IENLP VKVYV LNNQH LGHVY | | | | | |
| V22/1A | MGFL PAAAG ASYAN PGVTY VD1DG DGSFL MNVQE LAMIR IENLP VKVYV LNNQH LGHVY | | | | | |
| B73/7-4 | MGFL PAAAG ASYAN PGVTY VD1DG DGSFL MNVQE LAMIR IENLP VKVYV LNNQH LGHVY | | | | | |
| XI12/8A | MGFL PAAAG ASYAN PGVTY VD1DG DGSFL MNVQE LAMIR IENLP VKVYV LNNQH LGHVY | | | | | |
| | 550 | 560 | 570 | 580 | 590 | 600 |
| XI12/8A | QVEDR FYKAN RAHTY LGNPE NESEI YPDFV TIAKG FNIPA VRYTK KNEVR AAIKK MLETP | | | | | |
| V22/1A | QVEDR FYKAN RAHTY LGNPE NESEI YPDFV TIAKG FNIPA VRYTK KNEVR AAIKK MLETP | | | | | |
| B73/7-4 | QVEDR FYKAN RAHTY LGNPE NESEI YPDFV TIAKG FNIPA VRYTK KNEVR AAIKK MLETP | | | | | |
| XI12/8A | QVEDR FYKAN RAHTY LGNPE NESEI YPDFV TIAKG FNIPA VRYTK KNEVR AAIKK MLETP | | | | | |
| | 610 | 620 | 630 | | | |
| XI12/8A | GPYLL D11VP HQEHV LPHIP NGGAF KDMIL DGDGR TVY* | | | | | |
| | | | * | | | |
| V22/1A | GPYLL D11VP HQEHV LPHIP sGGAF KDMIL DGDGR TVY> | | | | | |
| B73/7-4 | GPYLL D11VP HQEHV LPHIP sGGAF KDMIL DGDGR TVY> | | | | | |
| XI12/8A | GPYLL D11VP HQEHV LPHIP NGGAF KDMIL DGDGR TVY | | | | | |

FIG.8C

(10)



Europäisches Patentamt
European Patent Office
Office européen des brevets



(11) Publication number:

0 525 384 A3

(12)

EUROPEAN PATENT APPLICATION

(21) Application number: 92110627.4

(22) Date of filing: 24.06.92

(51) Int. Cl.⁵: **C12N 15/60, C12N 9/88,
C12N 15/82, C12N 1/21,
C12N 5/10, A01H 5/00,
A01H 1/02**

(30) Priority: 31.07.91 US 737851

(41) Date of publication of application:
03.02.93 Bulletin 93/05

(54) Designated Contracting States:
**AT BE CH DE DK ES FR GB GR IT LI LU NL PT
SE**

(36) Date of deferred publication of the search report:
23.06.93 Bulletin 93/25

(71) Applicant: **AMERICAN CYANAMID COMPANY**
1937-West-Main Street P.O. Box 60
Stamford Connecticut 06904-0060(US)

(72) Inventor: **Dietrich, Gabriele Elfriede**
11 Merritt Lane
Rocky Hill, New Jersey 08543(US)

(74) Representative: **Wächtershäuser, Günter, Dr.**
Tal 29
W-8000 München 2 (DE)

(37) **Imidazolinone resistant ahas mutants.**

(37) The present invention relates to monocot genes encoding a mutant AHAS enzyme that is specifically resistant to imidazolinone herbicides. Exemplary of these genes are corn DNA sequences which encode an amino acid substitution at position 621 of the wild-type AHAS enzyme. The mutant gene can be used to transform other plants to herbicide resistance; in this regard, the invention also provides host cells and vectors containing the gene, which cells and vectors are useful in the transformation process.

EP 0 525 384 A3



European Patent
Office

EUROPEAN SEARCH REPORT

Application Number

EP 92 11 0627

Page 1

| DOCUMENTS CONSIDERED TO BE RELEVANT | | | |
|---|--|---|--|
| Category | Citation of document with indication, where appropriate, of relevant passages | Relevant to claim | CLASSIFICATION OF THE APPLICATION (Int. Cl.5) |
| X | WO-A-9 014 000 (ICI) 29 November 1990 * the whole document * | 4,10-12 | C12N15/60 C12N9/88 C12N15/82 C12N1/21 |
| D,X | EP-A-0 154 204 (MOLECULAR GENETICS) 11 September 1985 * the whole document * | 4,10-12 | C12N5/10 A01H5/00 A01H1/02 |
| X | CHEMICAL ABSTRACTS; vol. 112, 1990, Columbus, Ohio, US; abstract no. 193710, ANDERSON, P.C., ET AL. 'Herbicide tolerant mutants of corn' page 259 ; * abstract * & GENOME vol. 31, no. 2, 1989, pages 994 - 999 --- | 10,12 | --- |
| P,X | WO-A-9 208 794 (ICI) 29 May 1992 * the whole document * | 1-14 | --- |
| A | NUCLEIC ACIDS RESEARCH vol. 18, no. 8, 1990, ARLINGTON, VIRGINIA US page 2188 SATHASIVAN, K., ET AL. 'Nucleotide sequence of a mutant acetolactate synthase gene from an imidazolinone-resistant Arabidopsis thaliana var. Columbia' * the whole document * --- | 1-14 | C12N A01H |
| The present search report has been drawn up for all claims | | | |
| Place of search THE HAGUE | | Date of completion of the search 20 APRIL 1993 | Examiner MADDOX A.D. |
| CATEGORY OF CITED DOCUMENTS | | | |
| X : particularly relevant if taken alone Y : particularly relevant if combined with another document of the same category A : technological background O : non-written disclosure P : intermediate document I : theory or principle underlying the invention E : earlier patent document, but published on, or after the filing date D : document cited in the application L : document cited for other reasons * : member of the same patent family, corresponding document | | | |

EP0 FORM 100 (04/89)



European Patent
Office

EUROPEAN SEARCH REPORT

Application Number

EP 92 11 0627
Page 2

| DOCUMENTS CONSIDERED TO BE RELEVANT | | | |
|--|---|---|---|
| Category | Citation of document with indication, where appropriate, of relevant passages | Relevant to claim | CLASSIFICATION OF THE APPLICATION (Int. Cl.5) |
| A | PLANT PHYSIOLOGY SUPPLEMENT vol. 93, no. 1, May 1990, page 157 SATHASIVAN, K., ET AL. 'Isolation and characterization of mutant acetolactate synthase gene from an imidazolinone-resistant Arabidopsis thaliana var. Columbia' * abstract 917 | 1-14 | |
| A | EP-A-0 360 750 (CIBA-GEIGY) 28 March 1990 * examples 19,20 * | 1-14 | |
| | | | TECHNICAL FIELDS SEARCHED (Int. Cl.5) |
| The present search report has been drawn up for all claims | | | |
| Place of search THE HAGUE | | Date of completion of the search 20 APRIL 1993 | Examiner MADDOX A.D. |
| CATEGORY OF CITED DOCUMENTS | | | |
| <p>X : particularly relevant if taken alone Y : particularly relevant if combined with another document of the same category A : technological background O : non-written disclosure P : intermediate document</p> <p>T : theory or principle underlying the invention E : earlier patent document, but published on, or after the filing date D : document cited in the application L : document cited for other reasons</p> <p>a : member of the same patent family, corresponding document</p> | | | |

EP 92 11 0627 (P)